MS 08-382 Special Issue on Pharmaceuticals in the Environment, 28:12 Running Head: Secondary routes for human and ecological exposure to drugs Christian G. Daughton Chief, Environmental Chemistry Branch National Exposure Research Laboratory U.S. Environmental Protection Agency 944 East Harmon Avenue Las Vegas, NV 89119 USA (702) 798-2207 (702) 798-2142 (fax) daughton.christian@epa.gov Word count Text: 15,445 References: 4,459 Tables: 4,215 

Environmental Footprint of Pharmaceuticals: The Significance of Factors beyond Direct Excretion to Sewers Christian G. Daughton and Ilene Sue Ruhoy <sup>6</sup>Environmental Sciences Division, National Exposure Research Laboratory, Office of Research and Development, U.S. Environmental Protection Agency, 944 East Harmon Avenue, Las Vegas, Nevada 89119 \*Touro University Nevada, College of Osteopathic Medicine, Henderson, Nevada 89014, USA (Received 21 August 2008; Accepted 29 January 2009) 

Environmental Toxicology and Chemistry (SETAC).

\*To whom correspondence may be addressed (daughton.christian@epa.gov).

48

49

50

This ET&C Paper in Press manuscript is in its original unedited form and has not been

copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of

51

52

53

54

55

56

57

58

59

60

61

62

63

64

65

66

67

68

69

**Abstract**—The combined excretion of active pharmaceutical ingredients (APIs) via urine and feces is considered the primary route by which APIs from human pharmaceuticals enter the environment. Disposal of unwanted, leftover medications by flushing into sewers has been considered a secondary route – one that does not contribute substantially to overall environmental loadings. The present study presents the first comprehensive examination of secondary routes of API release to the environment and for direct but unintentional human exposure. These include: bathing, washing, and laundering, all of which release APIs remaining on skin from use of high-content dermal applications or from excretion to skin via sweating, and disposal of unused and partially used high-content devices. Also discussed are the health hazards associated with: partially used devices, medication disposal practices of consumers, and interpersonal dermal transfer of API residues. Understanding these secondary routes is important from the perspective of pollution prevention, as actions can be designed more easily for reducing the environmental impact of APIs compared with the route of direct excretion (via urine and feces), as well as for reducing the incidence of unintentional and purposeful poisonings of humans and pets and for improving the quality and cost-effectiveness of healthcare. Overall, unintentional exposure to APIs for humans via these routes is possibly more important than exposure to trace residues recycled from the environment in drinking water or foods.

**Keywords**–Pharmaceuticals: Excretion; Sweat; Poisoning; Disposal

### Introduction

70

71

72

73

74

75

76

77

78

79

80

81

82

83

84

85

86

87

88

89

90

91

Pharmaceuticals and personal care products (PPCPs) as environmental pollutants is a subject that has received exponentially growing attention since the late 1990s. The U.S. Environmental Protection Agency (U.S. EPA) maintains a large, publically available literature citation database for PPCPs ([1] http://www.epa.gov/ppcp/lit.html). It currently catalogs over 6,000 citations covering aspects that are directly or peripherally related to the entire spectrum of the risk paradigm – from origin and sources, to fate and transport, source control and waste treatment, ecological and human exposure, biological effects, pollution prevention, risk management, risk perception/communication, modeling, and others. Among these thousands of publications, however, fewer than 200 address any of the aspects of leftover (expired or unwanted) drugs and their disposal. None discuss the secondary routes by which active pharmaceutical ingredients (APIs) enter the environment (those beyond direct excretion to sewers) or that serve as source terms for modeling human exposure. The present study is the first comprehensive examination of the hazards of drug disposal and the potential significance of the secondary routes by which APIs enter the environment. This includes summarizing what the published literature covers as well as highlighting the data gaps and needs, and a framework, termed pharmEcokinetics (PEK), as the umbrella under which these processes and their relative significance might be better understood. Widespread occurrence of APIs in the environment is now well-established. Published reports of the occurrence of APIs in sewage, surface and groundwaters, sediments, sewage sludge, biota, and elsewhere in the environment total over 1,000 as of August 2008 ([1], http://www.epa.gov/ppcp/lit.html); many of these studies were catalyzed after the seminal 2002

This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).

publication of the initial nationwide monitoring study by the U.S. Geological Survey (USGS) [2].

The environmental presence of APIs is attributed primarily to raw or treated sewage (for human drugs) and to manure and lagoons (for veterinary drugs used in animal feeding operations); additional, less-obvious sources also exist, which can sometimes play important localized roles [3]. The major route by which APIs enter sewage is commonly accepted to be via urine and feces, with each contributing different relative amounts depending on the pharmacokinetics and structure of the individual API [4]. While other contributory routes, such as personal hygiene bathing or washing and the disposal of leftover medications by consumers, have been considered minor or inconsequential [5-7], no empirical evidence has yet been published to support this supposition.

Specifically with respect to the disposal route, prior work regarding leftover, unwanted medications has covered the following aspects: the broad spectrum of locales in society where unused drugs accumulate and from where they must be disposed or stockpiled [3, 8]; the many factors that lead to the accumulation of leftover medications, which then in turn eventually result in the need for their disposal [8]; the many approaches having the potential to minimize or reduce the accumulation of unused, leftover drugs and therefore reduce the need for disposal [8-10]; disposal of consumer drugs via collection programs in the U.S. [11]; the factors that encourage disposal to sewers versus other means of disposal such as trash or formal means of collection (e.g., take-back events) [12-14]; the first methodology by which accurate and comprehensive empirical data on the actual types and quantities of individual APIs that are disposed can be collected for a particular, defined population - namely coroner records from

decedent investigations [15]; and the human health, medical, and environmental ramifications and consequences of accumulated, leftover drugs [16, 17].

114

115

116

117

118

119

120

121

122

123

124

125

126

127

128

129

130

131

132

133

134

135

136

As for bathing or washing as a route of release and the hazards of leftover medications and the disposal process, no formal discussion has ever been presented to our knowledge, other than brief mentions ([3]).

In the present study, disposal is placed into a formal context for assessing its significance. The potential significance of washing and dermal transfer to other surfaces as contributory routes is also examined. Disposal and washing/bathing are discussed as the two most important secondary alternate routes of API release to the environment. The context required to assess their relative significance as contributory routes is developed. Understanding these routes is important from the perspective of pollution prevention, as actions can then be designed more easily for reducing their environmental impact compared with the route of direct excretion (via urine and feces), as well as for reducing the incidence of unintentional and purposeful poisonings of humans and pets. It is worth noting that while the literature on the larger topic of PPCPs as pollutants has grown dramatically since the mid-1990s [1], the total number of publications (beginning in the late 1980s) that tackle the questions surrounding disposal from a scientific perspective are few. This means that the single aspect of the problem involving environmental and human exposure having the greatest potential for control (i.e., disposal) has received the least attention. This bias has arisen perhaps because disposal has generally been assumed to contribute little to environmental residues compared with excretion.

Also introduced here is the concept of PEK. In simple terms, conventional pharmacokinetics (PK) deals with how a drug is processed in an organism (mainly, the time-course of drug concentration), with the prime focus being on the eventual concentration that

becomes bioavailable, so that therapeutic effect can be optimized and side-effects minimized:

"The activity of drugs in the body over a period of time, including the processes by which drugs are absorbed, distributed in the body, localized in the tissues, [transformed], and excreted" ([18], http://www.cancer.gov/templates/db\_alpha.aspx?CdrID=44324). Since PK focuses on the active (plasma) concentrations of drugs, excretion is only of indirect interest to pharmacologists (sometimes measured solely for mass balance and to get a better idea of the portion of API that might still be available within the body and to calculate half-lives). PharmEcokinetics is analogous to PK by also considering the fate of APIs in the environment (beginning at the point where an API or metabolite is excreted), with the primary difference being that its focus is decidedly not on plasma levels, but rather environmental levels, particularly wastewater, drinking water, and biosolids, as well as other locations (e.g., biota). The major aspects of PK that are of interest for PEK in environmental modeling are all of the pathways of excretion of unchanged parent API (and bioactive metabolites and labile conjugates).

To date, the routes of excretion that are factored into environmental modeling are urinary and fecal, both having been long assumed to be the major contributors to ambient environmental residues. While these generalizations are probably correct for APIs overall, the possibility has not been ruled out that they might not apply to certain APIs. While direct disposal of unused APIs is recognized as an additional source for entry to the environment, its significance is unknown and generally deemed to be inconsequential. Another route, which has essentially been ignored is release of API residues from the skin, by bathing and washing (including laundering of clothing and bedding contaminated by dermal contact) and by direct transfer via surface contact.

This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).

Disposal has two distinct contributors: leftover unused medications (i.e., expired, unwanted, or unused for other reasons), and partially used medications that retain residual API (delivery devices such as transdermal or transmucosal delivery systems). Approaches to pollution prevention could differ for these two sources. Washing, bathing, and dermal transfer by direct contact have three contributors: residues remaining after the administration of dermal medications - APIs for local treatment (topical use) and systemic treatment (transdermal/transmucosal delivery), residues remaining on the skin after removal of transdermal systems (e.g., patches), and residues excreted via sweat, a route that has only been briefly discussed with respect to its possible role in environmental pollution [16, 19].

Historically, consumers and other end users in the United States (U.S.) have disposed of leftover, unwanted medications by flushing them down sewer drains or by discarding them in the trash. For the vast majority of medications, the most prudent approach for addressing leftover medications is to avoid disposal to sewers. A number of countries (but not the U.S.) have long had programs where consumers can return leftover medications to pharmacies). In the last few years, various cities in the U.S. have begun implementing take-back collection programs, where consumers can return their unwanted medications [11]; in the U.S., however, these types of collection programs can be complicated by the presence of controlled substances, which can only be transferred by the prescription holder to law enforcement (and their deputies) or agents of the Drug Enforcement Administration (DEA), or among DEA registrants [3].

Drug diversion (the use of licit drugs for purposes that differ from their original purpose; recreational use is one example) is an important public health and safety concern and occurs by various routes, such as burglaries of residences and pharmacies, breaches of the manufacturing, distribution, prescribing, and dispensing chains, and theft by family and friends (e.g., teen

182

183

184

185

186

187

188

189

190

191

192

193

194

195

196

197

198

199

200

201

202

203

204

pharming) [20]. For medications that pose extraordinary and imminent hazards to humans (e.g., those subject to abuse or those having high acute toxicity), the possibility of unintentional poisonings or diversion for abuse must be minimized, as medications are a major cause of poisonings in the U.S. The United States Poison Control Centers recorded over 1,330,000 cases of unintentional non-fatal poisonings by chemicals in 2003, 42.6% involving children aged 5 and vounger. During 2001 to 2003, the United States Centers for Disease Control estimated over 53,000 children aged 4 and younger (72% of which were aged 1-2) suffered unintentional poisoning from OTC (over-the-counter) and prescription medications [21]. Nearly 10% required special medical care. Over 75% occurred in homes. A survey of death certificate data indicates a considerable presence of drug-related mortality, specifically overdoses [22]. Imprudent storage and disposal (e.g., to trash) is possibly a major cause of unintended exposures of those for whom the medication was not prescribed or intended, especially children [21]. Trade-offs are therefore required to best balance exposure of the environment (primarily via disposal to sewers) versus human exposure (e.g., via diversion from stored stockpiles of leftovers or from those medications disposed into the trash). It is widely accepted that a select number of medications are still best disposed by flushing to sewers as soon as they are no longer needed. This limited list of medications includes those that remain unused as well as certain ones (such as transdermal patches) that retain appreciable residuals after being completely or partially used. A list of these medications is highlighted by the White House Office of the National Drug Control Policy as part of their drug disposal guidance for consumers ([23], http://www.whitehousedrugpolicy.gov/publications/pdf/prescrip\_disposal.pdf). This guidance, however, is in a state of flux and is subject to modification, especially because of the intricacies presented by the Controlled Substances Act ([24],

http://www.deadiversion.usdoj.gov/fed\_regs/rules/2009/fr0121.htm). Note that unanticipated adverse ecological consequences have also occurred from the veterinary use of drugs that have resulted in drug-contaminated waste; two noteworthy examples are pentobarbital and diclofenac, which have resulted in considerable adverse impacts on populations of raptors and vultures [3].

A major unanswered question, however, is what portions of environmental residues of APIs originate as a result of intended therapeutic use, disposal, and washing/bathing. The approach presented here is intended for assessing the significance of these alternate pathways. The API contributions from these pathways is important in order to gage the possible efficacy of pollution prevention efforts for reducing environmental loads and to assess the hazard for humans. Evidence already exists that reducing the accumulation of drug waste has benefits with respect to healthcare, for which waste reduction is particularly attractive as a strategy because it imposes no limits on the overall usage of medications, it does not adversely affect the quality of healthcare [17], and may even serve to improve it [16].

In calculating predicted environmental concentrations, a variety of assumptions are required for the many inputs for a model's variables [25 - 27]. Three factors for which practically no empirical data are available are: the portions of medications that are disposed to sewers; the portions of APIs discharged to sewers resulting from the use of medications designed for dermal use; and the contributions from oral/parenteral formulations that are washed from the skin – as a result of excretion from eccrine and apocrine sweat.

With respect to disposal, the extensive examination of predicted environmental concentration calculations performed by Kostich and Lazorchak [25] had to assume that the following portions of medications are disposed to sewers: medicines prescribed for short-term therapy (15%), for long-term therapy (5%), and topical medicines (33%). Unfortunately, there

are no empirical data to validate these assumptions. These generalizations are likely much too high or too low when applied to many specific APIs.

Also with respect to disposal, another factor must be evaluated for delivery devices, especially transdermal systems, as the API residuals in used, high-content devices can be substantial (over 50% of the original API can remain); with patches, whose initial API content can exceed by 20 fold that which is eventually absorbed, more than 95% of the initial API content can remain in the used patch [28]. This means that for used patches, which are commonly disposed by flushing, the majority of the API from the used medication can be eventually disposed via sewers. Also, while the portion of API excreted via urine and feces is considered (as a major contributor), no consideration is given to the portion of unchanged API that might be excreted via sweat (because such data are extremely rare for prescription and OTC drugs) or the portion remaining on the skin after dermal application. Finally, the need to assume single, average values for individual factors for all APIs in general does not accommodate for the extreme ranges that actually exist among individual APIs.

Once a prescription drug is dispensed to an end user or an OTC medication is purchased, there are at least seven factors that have received little attention in previous modeling efforts and which determine whether disposal or bathing become important factors with respect to the overall occurrence of APIs in the aquatic environment (**Table 1**). **Figure 1** summarizes these factors and shows their interconnections.

In determining the significance of these secondary routes by which APIs can enter the environment, several key questions are prompted. For APIs with a presence in ambient waters, what individual portions or individual APIs originate from direct disposal of leftover, unwanted medications; release by bathing of residues remaining on the skin from dermally applied

medications; and release by bathing of residues remaining on the skin from excretion via sweat? Another source, related to the last two factors, is laundering, as drugs present on the skin can be transferred to clothing and bedding. Note that the release of topically applied drugs from domestic animals is also a source of APIs in the environment [29].

From these data, rankings could eventually be prepared to show which drugs contribute the most and least mass of APIs to the environment via disposal and via washing, in terms of absolute amounts as well as relative to the amounts contributed directly via excretion. This would allow the development and better targeting of pollution prevention measures and better targeted environmental monitoring.

Regardless of what percentage of APIs in the environment might be contributed to sewers by disposal or bathing, these practices could lead to transient, episodic spikes in API concentrations. These momentary concentrations could be orders of magnitude greater than what are being continually introduced via direct excretion [16]. Note, however, that those APIs for which disposal and bathing contribute the largest portion of their presence in ambient waters, this still would not reveal the relative importance of disposal or bathing with respect to the potential for actual adverse impact to the environment.

For a given API, the total mass discharged to sewers as a result of disposal and washing/bathing (WM) could be calculated from the factors in Table 1 according to:

269 
$$WM = UR(mass) \cdot [(DP \cdot PDs) + (SMP + RD + TD)]$$
 (1)

The relative significance (Sr) of the contribution from these alternate routes (versus direct excretion) for a particular API could be calculated by dividing the total mass contributed by disposal and washing/bathing by the total mass excreted unchanged in urine and feces:

 $Sr = WM \cdot [(UR - WM) \cdot PMP]^{-1}$  (2)

where UR is the usage rate; DP this is disposal potential (portion of API leftover); PDs are the method of disposal (portion disposed to sewers); PMP is the primary metabolic profile (portion excreted in urine/feces); SMP is the secondary metabolic profile (portion excreted by other routes such as sweat); RD is the route of delivery( portion remaining on skin after dermal application); and TD is the type of delivery system or container (portion of dose remaining in device).

As the portion of an API in sewage resulting from the alternative disposal routes increases (as WM increases, or as the portion used-as-intended decreases), Sr increases. As the portion resulting from disposal decreases, Sr approaches zero. In the absence of values for UR, however, Sr cannot be fully evaluated.

One factor regarding the correlation of disposal with intended usage needs to be emphasized, as it would complicate STET efforts at modeling. Sales are not linked in time to disposal. Disposal always occurs from sales made in the past. This time lag can also vary, forcing gross simplifications for the purposes of modeling. The significance of the time lag between dispensing and when leftovers will be disposed diminishes as the time period examined increases. In other words, the correlation over time between sales figures and if/when the drug is disposed will improve as the time period examined increases (perhaps extending out to the shelf-life of the drug). The consistency of correlation between sales and disposal over time is a function of the consistency in sales. For a medication whose usage (as reflected by sales) remains constant over time, the rate of disposal during a given time period will probably best correlate with the sales during that same time period even though the disposal results from sales during a prior time period. With this complication in mind, special circumstances become evident when

This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).

disposal could become the primary source of an API in the environment. For example, the environmental contributions from disposal might be greater than from intended usage if the sales for a medication dropped precipitously, for example, if the drug lost market share quickly or if it was recalled by the United States Food and Drug Administration. Likewise, the relative contributions from disposal would be less should a medication experience a rapid increase in sales, removed for example, an antiviral/antimicrobial drug dispensed during epidemics.

Clearly, to model the contributory origins of APIs present in the environment requires far more empirical data than currently exists. The empirical data necessary for many of the factors in Table 1 are not available. What is currently known regarding these previously unexplored factors is discussed in the remainder of the present study.

## Sweat as a route of excretion

While most unmetabolized, parent APIs are excreted via feces and urine, often overlooked is that measurable quantities of many APIs and/or their metabolites can be excreted via sweat. Excretion via sweat has been known at least since the 1950s, with one of the very early studies being published by Thaysen and Schwartz [30]. While the initial studies (up until the 1990s) focused on therapeutic drugs, interest has since shifted to illicit drugs, where sweat has become a matrix for monitoring illicit drug usage. Most of this literature therefore deals with what is known as sweat-patch testing as a means of non-invasive monitoring; see overviews by Rouen et al, ([31], http://notes.med.unsw.edu.au/ndarcweb.nsf/resources/TR\_18/\$file/TR.120.PDF) and Fortner [32]. Upon oral ingestion (or any other means of drug delivery), excretion of the original dose via sweat can continue for anywhere from a day to weeks. Excretion can begin in less than an hour, and varies from drug to drug. Excreted residues can then be collected on absorbent patches affixed to the skin. The concentration in sweat might vary depending on the sweating

rate, with the rate for some APIs remaining the same and others increasing with increasing rate of sweating [33].

319

320

321

322

323

324

325

326

327

328

329

330

331

332

333

334

335

336

337

338

339

340

Sweat contributes to two pathways for transporting APIs to the immediate and surrounding environments: direct exposure of others and contamination of surrounding objects via dermal contact (e.g., drugs released in sweat through the rows of pores located along fingerprint ridges are known to be deposited in fingerprints) [34], and release of APIs directly to sewers, via bathing and other hygiene activities.

Insensible sweat (passive diffusion through the skin) is produced at a rate of 300 to 700 ml/d per individual (at a rate of up to 100 g/m<sup>2</sup>/h in air temperatures below 31°C). With rigorous exercise, sensible sweat production (primarily from eccrine glands and secondarily from apocrine glands) can increase to 2 to 4 L/h for short periods or 1 L/h for prolonged periods. Eccrine sweat glands are distributed widely across the body, whereas the apocrine glands have very limited distribution; apocrine glands excrete via the hair follicles. As the largest organ of the body (and comprising 10% of body mass), the average skin surface area is very roughly 2 m<sup>2</sup> [35]. Drugs become incorporated with sweat via passive diffusion through cellular membranes driven by the concentration gradient established by the free drug in plasma and fat depots. Since sweat is normally very slightly acidic (pH <6.5) and blood is slightly alkaline, those drugs that are primarily non-ionized in plasma experience a negative concentration gradient across the skin (because they become ionized in the accumulating sweat). Therefore, excretion and accumulation in sweat favors those APIs that are neutral at around pH 7.4. See overviews by Tobin [36] and Fortner [32]. Excretion via sweat also seems to discriminate against polar metabolites. One extreme example is cocaine, which is extensively excreted in urine as ecgonine

This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).

methyl ester and its hydrolytic product, benzoylecgonine. But in sweat, cocaine is extensively excreted in its unchanged parent form [37].

While the concentrations of APIs in the aqueous natural environment are generally very low [7, 38], usually less than 1  $\mu$ g/L, it is not known what the relative contributions might be among urine/fecal excretion, disposal, or bathing. The extent and magnitude of excretion via sweat, and its significance with respect to contributing APIs to sewage, is clearly largely dependent on the amount (and type) of sweat that is generated per day (which can vary widely depending on the individual, the level of activity, level of hydration, the temperature/humidity, level of stress, the content and distribution across the skin surface of apocrine and eccrine sweat glands, and health and skin condition of the individual), the pH of the sweat, the plasma concentration and p $K_a$  of the API, and the bathing frequency, among other factors.

Despite the use of sweat-patch testing for illicit drugs, the published quantitative data on excretion of commercial drugs via sweat is rather limited. Most of the major studies are summarized in **Table 2**; note that in this discussion, sweat from eccrine and apocrine sources is not distinguished. The data on excretion via sweat is generally not obtained for the PK studies performed on a drug for registration purposes. Instead, these data are obtained during independent research studies. The excretion data provided by PK studies could perhaps be used, however, as an indirect indicator of the possible extent of excretion via sweat (e.g., by examining mass balance discrepancies of the percentage of an API not accounted for by excretion via urine and feces); but these studies are generally done under comfortable conditions where excretion via sweating would be minimized. If the excreted API is continually removed (e.g., via bathing or periodic re-contact with clothing), then the amount reabsorbed would be minimized. Also note that only recently has it become possible to accurately determine the actual concentration of

xenobiotics in sweat as determined with sweat-patch testing. Appenzeller et al. [39] normalized the quantities collected on patches to the content of sodium ion. Without normalization, excretion rates can only be roughly estimated by making assumptions regarding sweat volume production rate and collection efficiency.

Significance of excretion via sweat as a contributor to environmental residues

By performing some rough calculations, a general estimate of the relative contribution of APIs from sweat versus fecal/urine excretion can be obtained. The total amount of many APIs excreted via sweat may comprise very roughly up to 2 % of the total oral (or parenteral) dose. For those APIs that are the most extensively metabolized (e.g., when the percentage of parent API excreted unchanged and not in conjugated forms is less than a 2%), the contribution from sweat could prove to be an important factor.

The following serve as examples. Using the ratio of the areas of the sweat patch and body surface area, Pichini et al. [35] derived a crude estimate of the total amount of 3,4-methylenedioxymethamphetamine excreted via sweat 24 h after a 100-mg oral dose. The mean total mass excreted via sweat was estimated as 0.6 mg with an upper range of 1.5 mg (because of large inter-individual variability). This amounts to at least 0.6% (and 1.5%) of the total excreted (assuming methylenedioxymethamphetamine is extensively excreted unchanged).

After daily doses of 1,500 mg of ciprofloxacin, assuming a conservative rate of sweat production of 1 L/d, and assuming sweat concentrations ranging from 2 to 23  $\mu$ g/ml [40], the total daily excretion of ciprofloxacin in sweat would range from 2 mg/d (2  $\mu$ g/ml • 1000 ml/d) to 23 mg/d. Assuming that ciprofloxacin is extensively excreted unchanged, the fraction excreted via sweat would be roughly 0.1 to 1.5% of the total available.

This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).

For drugs that are extensively excreted unchanged, the portions contributed by sweat are measurable but, as seen from these examples, not very significant (e.g., roughly less than 2%). For these drugs, the importance of excretion via sweat would more likely be dispersion to the immediate environment via dermal contact. For drugs that are extensively metabolized, however, the contribution to excreted APIs via sweat could be considerable and should probably be considered in predictive fate models.

Consider another example applied to a drug (fentanyl) that is more extensively metabolized (less than 8% excreted unchanged in the urine). In this case, the majority of the dose that is excreted might be excreted via sweat rather than urine. Schneider et al. [41] calculated the amount of fentanyl excreted via sweat as ranging from 19 to 150 µg/d, which translates to 3 to 25% of the total daily dose (600 µg). Relative to the amount imputed to be excreted in the urine (8%), the relative amount contributed by sweat would have ranged from 40 to 300%. As for many drugs, measurable quantities are also excreted via the hair (through apocrine sweat), but this is harder to quantify on a per-body basis. So for a drug that is not extensively excreted in the urine or feces unchanged, the portion excreted via sweat could be comparatively significant.

Using the data of Schneider et al. [41], a series of calculations were made using a different approach. The minimum mass of fentanyl excreted per day via sweat could be calculated from the minimum amount found on a sweat patch (5.7 ng) and using  $1.5 \text{ m}^2$  as the body skin surface area. The maximum mass of fentanyl excreted per day via sweat could be calculated from the maximum amount found on a patch (88 ng) and using  $2.0 \text{ m}^2$  as the body skin surface area. Calculating the number of patches that could hypothetically cover the body (total area in mm<sup>2</sup> divided by  $1480 \text{ mm}^2/\text{patch}$ ), and using the fraction of a day during which sweat was collected (0.42 d = 611 mins), the range of fentanyl mass excreted via sweat would have been 17 to 284 min

This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).

μg/d/body. Since the presumed daily dose was 600 μg, the percentage of the dose excreted via sweat could have ranged from 2.8 to 47%. It must be noted, however, that the heterogeneity of sweat excretion (as well as the concentrations of APIs in different microenvironments of sweat) could vary greatly. Therefore, all extrapolations on total amounts of API excreted are subject to considerable error. Insufficient data exist regarding API excretion to fully understand total-body excretion via sweat.

Assuming that only 8% of a fentanyl dose is excreted unchanged via urine each day (which in this example is 48  $\mu$ g), the range in mass excreted via sweat would have been equivalent to that excreted from the following number of daily doses delivered via patch (in terms of relative contributions of fentanyl to sewage): 17/48 to 284/48 = 0.35 to 5.9. Of course, there are many variables, including actual skin area over which sweating occurs, uniformity of sweating over the body, uniformity of sweating rate (which could be seen as varying over a 15-fold range), uniformity of excretion via sweat, uniformity of rate of fentanyl absorption (which determines plasma concentration), etc. But from these crude calculations, the contribution of fentanyl to sewers via washing of sweat from the body could be equivalent to 30 to 600% of the mass originating from urine, in agreement with the estimates provided by Schneider et al. [41].

With respect to medications, appropriate exposure results from use by those for whom the API was intended and for whom the API was deemed safe, and inappropriate exposure occurs to those for whom the API was not intended (or for whom the API is contraindicated, or for whom the exposure was unwelcome). Inappropriate exposure to APIs via interpersonal dermal transfer (or hand-mouth contact) might prove to be a more important source of exposure than exposure via drinking water. A comparative yardstick might be that the occurrence of APIs in sweat can reach concentrations at least 3 orders of magnitude higher than those eventually occurring from

recycling of residues from the environment via drinking water. Drinking water concentrations are generally much less than 1 μg/L [42] versus concentrations in sweat, which are roughly 1 μg/ml and higher. The very limited numbers of oral/parenteral APIs that eventually make their way into finished drinking water [42] must survive a series of steps that successively reduce their concentrations, including absorption and metabolism, sewage treatment, dilution in receiving water, environmental transformation, sorption to sediments, and final polishing to produce finished drinking water. A broader spectrum of APIs at much higher concentrations could therefore occur in sweat, including those that are otherwise extensively metabolized.

While direct exposure to APIs via contact with the sweat of others has unknown significance, the APIs excreted from the skin of those taking medications (including those undergoing chemotherapy) have the potential to be fully released from the entire body in public spas and swimming pools. This is a scenario where inappropriate or unwanted dermal contact could occur to concentrations higher than in waters from the ambient environment (e.g., > 1 ppb, ng/L). For those undergoing polypharmacy, the release of multiple APIs would likely occur.

The use of recently developed ambient surface-sampling/direct desorption mass spectrometry techniques (such as DESI, DART, and DAPCI or single-particle aerosol mass spectrometry – SPAMS) for the very fast in vivo surface-analysis of tissues could prove very useful for the broad survey of the prevalence of APIs excreted to the surface of skin, as well as items commonly touched by the public. These techniques excel at rapid identification of chemicals sorbed to complex solid substrates. The abilities of these techniques to readily detect drugs and metabolites on skin have been demonstrated by Martin et al. [43], Takats et al. [44], and Williams et al. [45]. Application of this type of technique could be used to quickly reveal the

extent and magnitude of drug excretion via skin and the indirect contamination by APIs by dermal transfer.

A final note is warranted regarding the significance of excretion via sweat. Much has been published regarding the growing prevalence of antibiotic resistance, especially among human pathogens. Excretion of antibiotics via sweat has been proposed as a possible major means of quickly promoting and spreading resistance. The comparatively higher and sustained concentrations on skin can serve to expose dermal bacteria, which can then be readily transferred to other locations or people. This has been demonstrated by Høiby and others [40, 46], who documented the excretion onto skin of floxacin and β-lactam antibiotics where bacteria would come into ready contact. This could be an overlooked cause of transmission of multiresistance among bacteria in hospitals and other care facilities that routinely administer antibiotics.

### Chemotherapeutics in sweat

Excretion of chemotherapeutics via sweat is well established, but its overall significance as a secondary exposure route for others is not. That chemotherapeutics are excreted via sweat is reflected by its becoming recognized as a primary cause of a variety of adverse cutaneous effects during chemotherapy (e.g., doxorubicin), including hand-foot syndrome (hand-foot skin reaction) [47, 48] and hyperpigmentation and alterations to nails. The specific formulation can enhance the excretion of the API via sweat.

But with respect to unanticipated exposure, this route of excretion holds the potential for promoting subsequent incidental exposures for others and poses higher risks than for other drugs because of the extreme cytotoxicity and mutagenicity of oncolytics. Excretion via sweat

undoubtedly also plays a role in the development of hypersensitivity to certain other drugs since it ensures skin contact with drugs not intended for dermal application.

Early studies indirectly measured the excretion of chemotherapeutics via sweat by mutagenicity assays. For example, a 1988 study showed that sweat collected from patients treated with cyclophosphamide and other antineoplastics showed greater mutagenicity than controls 8 h after treatment [49]. A mean concentration of methotrexate in sweat was measured as 725 ng/ml (mean maximal concentration of 1.7 μg/ml), calculated as translating into excretion of 300 μg per day through sweat [50]. Other studies provide strong indirect evidence that sweat conveys chemotherapeutics outside the body. These studies have focused on studies of occupational exposure [51], where bedding becomes contaminated and serves as a route of exposure for healthcare workers and especially those working outside hospitals, such as home care providers [52]; workers in laundry facilities were noted as having the potential for higher exposures to antineoplastics than oncology nurses during the handling of bed sheets.

### Chemotherapeutics and pulmonary exposure

Occupational exposure to antineoplastic agents has been well documented, especially direct exposure from the compounding, preparation, administration, and disposal of these highly toxic chemicals. Of the many routes of exposure, however, the excretion of residues via sweat (and breathing) of patients has been less understood. Several chemotherapeutics have appreciable vapor pressures. These include: carmustine, cyclophosphamide, ifosfamide, thiotepa, and mustargen [53]. Others have much lower vapor pressures: doxorubicin, cisplatin, etoposide, 5-fluorouracil, and mitomycin. Kiffmeyer et al. [54] determined that the vapor pressures of five antineoplastics (carmustine, cisplatin, cyclophosphamide, etoposide and fluorouracil; and one

antimicrobial drug, fosfomycin) were all low but with carmustine having a vapor pressure one order of magnitude higher. Nevertheless, cyclophosphamide was still detected in the gas phase in 7 out of 15 locations, at levels ranging from 45 ng/m³ to  $13\mu g/m³$ . Inhalation of excreted cytotoxics could also be enhanced for those who work in laundry facilities that clean bedding and clothes from patients [52], although Fransman et al. [55] did not detect vaporization of antineoplastics from bedding at a laundry facility.

These data also indicate the theoretical potential for pulmonary exposure to the expired breath from those undergoing treatment. Fransman ([56], http://igitur-archive.library.uu.nl/dissertations/2006-1003-200854/full.pdf#page=131) notes that exposure to people or animals associating with those undergoing treatment with antineoplastic drugs has not been investigated. Unintended exposure in these settings could prove important with the increasing usage of antineoplastics in outpatient and veterinary clinics and since more intimate

### Dermal application

and chronic interpersonal contact can occur in the household.

The continuing trend toward the dermal application of drugs will increase the probability of drugs being introduced to the environment as a result of: release via bathing of concentrated residues remaining on skin and discarding the used delivery device (e.g., patches) which often contains very high levels of residues (sometimes considerably greater amounts than would have been needed orally). It could also increase the unintended risk of exposure to others by direct dermal-dermal contact and transfer, and from indirect exposure via contact with contaminated objects.

This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).

521

522

523

524

525

526

527

528

529

530

531

532

533

534

535

536

537

538

539

540

541

542

A wide array of drugs are available in topical form ([57], http://formulary.prescribingreference.com/dermatological disorders; [58]). See Table 3 for those that are commonly employed. The concentrations of APIs in these topical preparations range from a fraction of a percent to 5% and more, by weight. They include potent steroids, antibiotics, pesticides (e.g., lindane, malathion), immunomodulators (e.g., pimecrolimus), a psychotropic (doxepin), and cytotoxics (e.g., fluorouracil). Some of these drugs have no routine oral use (because of toxicity or facile metabolism), such as tolnaftate, ciclopirox, flurandrenolide, and imiguimod. For these drugs, bathing (and disposal) is most likely to account for the vast majority of any residues that might be detected in the environment. For others that also have equivalent oral uses, but are extensively metabolized (little excretion of unchanged API), bathing could still be a major contributor of residues to sewage. Those APIs with equivalent dermal and oral uses, but which are extensively excreted unchanged, are highlighted in Table 3. This group comprises the only topical APIs where washing and bathing could be competing with excretion from oral/parenteral use in terms of contribution to the environment and therefore where bathing would be a less important source. The APIs in this group (highlighted by footnotes in Table 3) are: acyclorvir, doxepin, fluorouracil, metronidazole, neomycin, nystatin, polymyxin, sulfadiazine, tobramycin, and tretinoin. All the remaining APIs in Table 3 (those not highlighted), if detected in the environment, could have origins primarily from dermal application. These latter APIs could be ranked according to overall usage rates (e.g., total mass sold) and potency to guide the selection of those to include in targeted monitoring in order to gage their potential frequency and extent of occurrence in the environment. In terms of accounting for bathing as a source term in fate

This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).

models, another source of APIs on the skin to account for is the residue remaining after the removal of a transdermal device such as a patch.

Among those topical/transdermal medications with no oral equivalents and which also have minimal excretion (of the absorbed dose), those sharing common mechanisms or modes of action (and for which dose addition might therefore be an important exposure consideration) could prove to be the most important with respect to environmental hazard. The corticosteroids, for example, all affect the hypothalamus-pituitary-adrenal axis, especially those that are not approved for oral or parenteral use. Many of the antibiotics could promote the selection for antibiotic resistance on the surface of skin since their localized concentrations can be extremely high [40, 46].

Comparing the mass of API residue remaining in a used delivery device or the residue remaining on the skin, with the mass that would be excreted if the API has been taken orally (or endogenously produced, such as certain hormones), can provide insight as to the relative significance of the pathways. For example, one recently introduced formulation is a metered-dose transdermal spray of estradiol (EvaMist, Vivus) where each metered dose (containing 1.7% estradiol) delivers 1.53 mg of 17β-estradiol. An estradiol gel (Estrogel, Solvay Pharmaceuticals) contains 0.06% estradiol, and a 1.25-g dose of the formulated gel contains 750 μg. The various reference ranges for urinary excretion of endogenous estradiol (assuming no deconjugation, which can be substantial, [59]) range from 10 to 100 μg/d (depending on the woman's age and health), or up to 30 mg/d (during pregnancy) [60]. Assuming a dermal estradiol absorption efficiency of 17% (24-h absorption reported for Estrogel [61]), one dose of the spray or gel could leave on the skin 1.3 mg or 0.6 mg of estradiol, roughly the endogenous amount excreted daily by 6 to 130 women who are not pregnant.

This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).

Another example is testosterone. A high-content gel form of testosterone (Androgel, 1% testosterone) has a maximum daily dermal dose of 100 mg of testosterone. Approximately 10% is systemically absorbed. Assuming the remainder (90 mg) is eventually washed from the skin, and assuming that the combined urinary excretion of free and conjugated endogenous testosterone from adult males ranges up to 0.3 mg/d (calculated from Al-Dujaili [62] and Timón Andrada et al. [63]), the daily use of testosterone gel could contribute a mass of testosterone equivalent to that excreted naturally from 300 (90/0.3) males.

An example of a dermal drug that also has oral formulations is ketoconazole. Once absorbed, only a fraction of a percent is excreted unchanged, meaning that except for the unabsorbed oral dose, bathing (and disposal) could also be the major source of this API in the environment. To assess the significance of dermal drugs as a contributory route to the environment, the following data would need to be compiled for each: fraction of dermal API not absorbed across the dermis (or residue left on skin after removal of a transdermal device), fraction of oral form not absorbed from the gut, and fraction of API excreted unchanged (as well as in easily hydrolyzable conjugates).

### Pollution reduction

Possible approaches that might help to reduce the introduction of dermal APIs to sewers prior to bathing include: removal of as much of the product from the skin as possible with an absorbent wipe such as toilet paper or cotton balls and then disposing in the trash; for preparations that have dried on the skin (such as gels), adding an oil (such as olive oil or hand cream) to the wipe might enhance removal; development of hand dispensers for topical drugs that minimize over-application (too large a quantity and/or applied over too large an area), which is difficult to avoid with many topical formulations; development of hand dispensers that permit more accurate dispensing to the target site with minimal wastage or over-spreading; and formulations that improve transdermal flux (which would also allow lower applied doses).

Overviews of current and future transdermal systems and technologies are provided by Wilkosz and Bogner [28] and Tanner and Marks [64].

### APIs commonly used in topical medications

The APIs commonly used in topical medications (excluding drugs delivered by transdermal systems) are listed in Table 3. Except where noted, these are the APIs for which the potential is highest that dermal application (as opposed to excretion) is a source for environmental residues. Some of these APIs are also used in oral and parenteral medications. Annotated in Table 3 is a rough categorization of the portion of an API that can be excreted unchanged. Those remaining APIs in the Table that cannot be extensively excreted, if detected in the environment, would have the higher possibility of having originated from bathing (as opposed to excretion via urine or feces). Of this sub-group of topical APIs, data from environmental monitoring exist only for a select few; these data are compiled in Table 4.

While the existing data show these APIs present in waters at sub-µg/L concentrations (except for crotamiton), those that belong to the same therapeutic class (such as the corticosteroids or antibiotics) have the potential for combined action via concentration (or dose) addition. Little is known regarding the environmental occurrence of the corticosteroids, as the first papers appeared only recently [65-67]. Note that clotrimazole is included on the List of Chemicals for Priority Action by the Convention by the OSPAR Commission for the Protection of the Marine Environment of the North-east Atlantic ([68], http://www.ospar.org/documents/dbase/decrecs/agreements/04-12e\_List%20of%20Chemicals%20for%20Priority%20action.doc). Also note that both clotrimazole and terbinafine were identified using a QSAR approach as among the top 10 chemical substances targeted for further screening ([69], http://www.environment-agency.gov.uk/commondata/acrobat/p601206trv2\_578719.pdf.)

Biopharmaceutics Drug Disposition Classification System

The Biopharmaceutics Classification System (BCS), developed by Amidon et al. [70], is a system for classifying APIs according to bioavailability. The BCS essentially categorizes APIs that are orally administered according to the four combinations of permeability and solubility, because absorption largely depends on solubilization of an API across the intestine. Wu and Benet [71] transformed this system to the Biopharmaceutics Drug Disposition Classification System (BDDCS), which categorizes APIs for oral administration according to the four combinations of solubility and metabolism. The BDDCS categories 1 and 2 are subject to a wide array of metabolic pathways leading to extensive excretion of metabolites, whereas categories 3 and 4 primarily are poorly metabolized and therefore are eliminated unchanged in the urine and

This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).

bile. A wide spectrum of other variables, such as food intake and its composition, also affect excretion [72]. Wu and Benet [71] state that very few APIs undergo intermediate metabolism (e.g., 50%). They are either extensively metabolized or largely excreted unchanged.

628

629

630

631

632

633

634

635

636

637

638

639

640

641

642

643

644

645

646

647

648

649

650

The BDDCS categories 1 and 2 are of interest with respect to understanding the significance of APIs that are administered topically as a primary source for the API in the environment. The environmental presence of those topical APIs that do not have an oral or parenteral equivalent will clearly be a direct function of the extent of their intended usage, which then leads to introduction to sewage via bathing. But for those topical drugs that also have oral or parenteral uses, the significance of the topical use will be a function of whether oral and parenteral use is accompanied by extensive metabolism and therefore little excretion of the unchanged parent API. Therefore, APIs in BDDCS categories 1 and 2 will contribute little unchanged parent API to the environment via excretion by urine and feces. The dermal use of these APIs could be responsible for the largest portion of the parent APIs in the environment. Since a trend is emerging for new molecular entities to be highly permeable, poorly soluble, extensively metabolized compounds (BDDCS Class 2) [72], this means that for those new molecular entities designed for dermal and oral/parenteral use, bathing and washing could play increasingly important roles with respect to release of APIs to the environment. This also means that disposal to sewers would have the potential to also grow in importance as a source, if newer drugs will be extensively metabolized. Note that of the dermal APIs listed in Table 4 and that also have oral use, four are listed in categories 3 and 4 by Wu and Benet [71]: acyclovir, neomycin, nystatin, and erythromycin. The primary source for APIs in the environment from these classes will probably continue to be direct excretion.

This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).

## Interpersonal dermal transfer

Considerable residues of APIs on skin (from dermal excretion, from topical application, and remaining after removal of transdermal systems) have the potential for being transferred directly to other persons or to surfaces with which others come into contact. Just as with exposure to APIs via drinking water, for most people this would constitute unexpected, unwelcomed exposure [42].

While direct dermal-dermal contact obviously increases the probability of interpersonal passive transfer, the possibility also exists for indirect human exposure during daily routine activities via dermal contact with surfaces previously contaminated with APIs from dermal products that remain on the hands of those who have personally applied topical drugs. One noteworthy example is hormonal preparations, such as testosterone, progesterone, and estradiol – those that are dermally applied as preparations containing very high concentrations (percent levels) by rubbing onto the skin with fingers or the hand. Even after hand washing, substantial residues can remain, resulting in physiologically significant exposures for others. Indirect transfer might be possible by contamination of inanimate objects (e.g., door handles, telephones, keyboards, plumbing fixtures, clothing, currency, etc.) followed with contact by others. The APIs from all types of medications applied by hand or from devices that are touched (e.g., new and used medicinal patches) clearly have the potential for widespread dispersion by these means.

The propensity for topical medications to get dispersed beyond their application sites was demonstrated over 20 years ago with the use of tetracycline. When topically applied, tetracycline was demonstrated to not remain in its original location, but rather to be transferred to other parts of the body. The degree and pattern of transfer was a function of the original site of application, the individual patient, and especially the vehicle in which the tetracycline was prepared (i.e.,

This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).

ointment, cream, lotion, or tincture) [73]. This demonstrated the ease with which dermal applications could be transferred.

Several cases have been reported of incidental, passive dermal transfer from parents to children. For example, androgenic steroids such as testosterone (cream) [74-77] and 4-androstenediol [74] topically applied to adults have resulted in profound physiological changes in children (substantial virilization of boys and girls) after interpersonal dermal contact. The vehicle in which the testosterone is delivered could play a role in the potential for transfer, as an alcoholic gel preparation seemed to prevent even purposeful interpersonal transfer [78]; the same was noted for a gel-form of estradiol [79]. But appreciable transfer resulting from a different gel formulation of testosterone was evident in another study, probably from much longer chronic contact [80]. One hour after dermal application of estradiol, purposeful interpersonal skin contact (for 15 mins), resulted in measurable systemic uptake by a naive recipient, largely because the majority of the initially applied dose remained on the skin surface for extended periods [81].

The ease with which dermally applied drugs can be transferred by contact, and their sustained persistence on the skin even after repeated washing, is shown by the contamination that can be introduced to laboratory analyses. After applying 5% progesterone cream by fingers to the body, simply transferring a sample with a pipet introduced considerable background levels of progesterone, even after the fifth hand washing. Contamination could even occur when using gloves, simply by pulling the gloves from their storage box by gripping a finger tip [82]. The same problem has also been noted for a technician who had been using topical testosterone (in gel form) and performing tests for testosterone. Contamination on the fingers led to very high errant test results [83]; the authors noted that it was not possible to remove all traces from the fingers. Also worth noting is that since residues of those drugs that are topically applied (usually

in large quantities) can remain on the skin, and since many APIs are excreted through the skin, unique challenges are posed for environmental monitoring. Stringent quality control measures must be implemented to guard against contamination during sampling, monitoring, and sample preparation; a comprehensive system of blanks is particularly important.

# Residuals remaining in used delivery systems

Although the topic of drug disposal primarily concerns leftover medications, completely used and partially used medications (especially non-oral delivery systems or devices) also serve as a source of APIs during disposal, as the remaining residuals in their leftover contents can represent a substantial portion of the amount present in new, unused devices. Leftover residuals in delivery devices is an issue only recently suggested as a source term needing further evaluation [3, 84]. This aspect has not been accounted for in source terms for fate models, and would be quite difficult to accommodate in a realistic manner, as the residual quantities would vary immensely depending on the type of device, its duration of use, and patient compliance.

These used devices themselves can also serve as a considerable acute hazard, as they are responsible for documented morbidity and mortality due to poisonings from unintended exposure and abuse. As a prime example, consider the list of APIs used most commonly in patch delivery devices designed to administer sustained dermal doses (Table 5). This table also shows the mass content per device, a rough estimate of the number of lethal oral doses in an unused device, and a rough estimate of the equivalent number of oral doses required to contribute the same mass of API if the unused device were flushed to the sewer. A major variable in determining the quantity of residual is the conditions under which the device is used. Failure to clean or dry the skin prior to application, for example, impedes permeation of the dermal layer.

Of the drugs listed, fentanyl patches (either new or used) have an API content sufficient for roughly up to 10 lethal oral doses in adults. Clonidine, nicotine, and possibly lidocaine patches have roughly sufficient API for multiple lethal doses in children or pets. These patches clearly require special care to ensure fast, secure, and safe disposal. These instances are not reflective the much higher possible incidence of morbidity from exposures to other APIs or lower doses.

By using the daily doses and excretion efficiencies for the oral versions, the quantity of an API released to sewers from disposal of a device can be compared with the API released from oral doses. Several devices, if disposed to sewers unused, would contribute the equivalent of thousands of oral doses (after accounting for pharmacokinetic data for excretion of unchanged API): methylphenidate (equivalent to 3,280 oral doses), rivastigmine (1,200), and nitroglycerin (2,667). Others would contribute the equivalent of hundreds of oral doses: clonidine (188), ethynylestradiol (214), oxybutynin (720), norethindrone (192), and norelgestromin (240). The residual content of used patches was available only for fentanyl, where a used patch would still be equivalent to hundreds of oral doses (420) [85]. Some patches would serve as unique contributors to the environment since oral equivalents of their APIs do not exist; these include rotigotine, flurandrenolide, and lidocaine.

The residual APIs in transdermal therapeutic systems (or other drug delivery devices designed for external extended release) can represent a substantial portion of the amount present in new or unused devices. It can also be considerable when compared with oral daily doses. This pertains especially to APIs formulated in transdermal and transmucosal devices. For transdermal patches, as an example, the residual is a function of how efficiently the API is absorbed across the skin and how long the patch is left in place. This adds an important but highly variable dimension to calculating the significance of disposal compared with excretion. The amount of API that is

This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).

retained on the skin surface from transdermal delivery devices can also be appreciable, as interdermal transfer of clinically significant amounts from these particularly concentrated areas can then occur between individuals as a result of bodily contact. This has been documented, for example, for estradiol [81], where the authors noted that "clinically significant transfer of topical bioactive drugs can occur."

A major concern regarding these devices (as with all APIs reformulated for low-dose extended release) is the purposeful circumventing of the design in order to acquire a high-dose immediate-release drug that can be taken via a direct route (such as by mouth, nose, or intravenous); this is an approach used by drug abusers. Indeed, design of devices to accommodate new delivery forms of APIs already in therapeutic use can lead to diversion and abuse – because of their high content of API [86]. Some APIs pose extreme risks and are tightly controlled under formal restricted-access programs, which impose restrictions on various aspects of prescribing, dispensing, or patient usage in order to reduce the risk of diversion, abuse, and imprudent use [87]. These risks largely fall into three major categories: potential for abuse (e.g., buprenorphine), severe adverse drug reactions (e.g., clozapine), and teratogenicity (e.g., thalidomide, isotretinoin). The latter are two examples of those for which inadvertent exposure must be minimized.

The first transdermal therapeutic system was a transdermal patch (incorporating scopolamine for motion sickness), approved by the United States Food and Drug Administration in 1979. This was followed by the development of the nicotine patch. An overview of transdermal systems is provided [88]; the number of therapeutic classes being formulated for dermal transfer continually expands, now including such drugs as psychotropics. Most drugs administered by patch pose four main hazards: potential for abuse of used patches, which can contain acutely toxic residual doses when administered by alternative routes (e.g., oral ingestion), potential for accidental poisoning

by used (as well as new) patches, which can be accidentally ingested by infants, toddlers, and pets, when disposed to sewers, used patches can contribute a mass of API equivalent to that resulting from excretion from multiple doses of oral formulations, and residues remaining on the skin after a patch is removed can be substantial, contributing to API release to sewers during bathing or to transfer to others by inter-dermal contact.

Delivery devices that usually are not disposed by flushing can also pose eventual exposure hazards, especially if disposed to trash that is landfilled [89]. As one example, Guerts et al. [90] calculated that the ethynylestradiol (EE2) remaining in used vaginal contraceptive rings is roughly 85% of the initial amount, corresponding to roughly 2.4 mg of EE2. This EE2 is then available for accumulating in landfill leachate; the same concern would apply to EE2 implants once removed by a physician. Ethynylestradiol is an extremely potent endocrine disruptor in the aquatic environment, having profound effects in fish populations at concentrations in the low parts-per-trillion range [91].

The acute risks posed by used delivery systems containing substantial API residues have been amply demonstrated. Unintentional poisonings and abuse are not uncommon. After 3 d of use, fentanyl patches have been reported to retain 28 to 84% of their original fentanyl content, more than sufficient for a lethal oral dose should the patch be applied dermally on an opioid-naive person or be ingested, for example, by an infant [85]. Note that a new 2.5-mg fentanyl patch contains the equivalent of about twelve 200-µg fentanyl lozenges (which are available in formulations of 200 to 1,600 µg in 200-µg increments). The residue in a used 2.5-mg patch might be equivalent to four to ten 200-µg lozenges.

Patches are indeed a known cause of fatal poisonings [92] after intentional ingestion by adults [93] and by children [94, 95], as well as by injection of API extracted from patches [96]

This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).

790

791

792

793

794

795

796

797

798

799

800

801

802

803

804

805

806

807

808

809

810

and by misuse and abuse of patches by misapplication and inappropriate application in numerous different ways [97]. Used nicotine patches have been ingested and applied dermally by children [98]. Used patches may be more likely than new medications to be accessed by children as they can be forgotten once removed.

A continuing trend toward designing new and existing APIs in dermal delivery systems could serve to reduce the ameliorative role that metabolism would normally play in reducing the load of APIs in the environment; the API content of dermal delivery systems is also often much larger than required for oral doses. This trend could increase the significance of used and unused leftover medications as a source of APIs in the environment, particularly for those APIs that would otherwise be extensively metabolized if consumed orally or parenterally. The unused portion of APIs in delivery devices clearly serves as a reservoir of APIs that may require additional attention with regard to disposal. While flushing might currently be the best alternative for quickly ensuring that used patches are not accessible to others, there are situations where flushing non-soluble materials is problematic, such as with septic systems. Because of its very high potential for abuse or accidental poisonings, used fentanyl patches pose exceptional challenges, as even patches on decedents are known to be diverted and reused [99] and have led to overdose and death [100]. Except for patches, most devices have very low potential for disposal via flushing (inhalers, venipuncture and other injection devices), so their API residues are not prone to immediately entering sewage. One way to reduce the significance of residual APIs remaining in transdermal and topical (e.g., cream and gel) applications is to improve the efficiency of dermal permeation/absorption (via reformulation); this would also allow the use of less API per application, thereby reducing the remainder yet further.

With respect to veterinary practice, the use of medicated feeds can contain substantial concentrations of drugs such as hormonal growth promoters and antibiotics. Unused feed and feed incompletely consumed (e.g., in aquaculture, where large portions sink before being consumed) can contribute residues to the environment [101] or be eaten by non-target animals.

### Residuals and the hazards of attempted medication destruction

The desire for methods that consumers can use to render unwanted, leftover medications unusable (prior to disposal) has led to recommendations to alter the physical form of the medication. One example is the SMARxT disposal campaign, which advises: "Pour medication into a sealable plastic bag. If medication is a solid (pill, liquid capsule, etc.), crush it or add water to dissolve it" or "add kitty litter, sawdust, coffee grounds,"([102], http://www.smarxtdisposal.net). But guidance aimed at altering or destroying medications poses acute hazards for people and pets and also possibly facilitates the entry of APIs to the environment.

Practices that attempt to render medications unusable by physical alteration not only do not prevent diversion to drug abusers – who can easily reclaim the APIs – they also pose additional risks. The need for new approaches for safe and environmentally prudent drug disposal has been discussed [8]. The magnitude of leftover medications has been documented in a number of publications, a recent one being De Bolle, et al [103].

A form of medication alteration long used in medical care is a common practice known as dose-form modification, used especially in long-term care facilities where patients often refuse oral medication or have difficulty swallowing (dysphagia) [104]. Healthcare professionals often resort to dose-form modification to get these patients into compliance. This commonly involves

This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).

crushing pills or opening capsules and transferring to a more easily administered format (e.g., mixing in sweetened food). Dose-form modification is controversial because it has the potential to radically alter the pharmacokinetics of medications that are specially formulated to release APIs gradually (e.g., special-release medication forms such as extended release and delayed release formulations). Physical modification can greatly reduce the time required for an API to reach a maximum plasma concentration, and this concentration can often exceed the threshold for adverse effects, because all of the API is released at once instead of over an extended time (e.g., a 12- or 24-h dose delivered all at once). It is widely recognized by drug abusers that pill crushing can lead to greatly enhanced biological activity [105].

Once crushed, the design of extended release tablets is defeated, making immediately bioavailable their entire contents of APIs. Those opiate medications containing APIs that are extremely potent can contain up to several lethal doses per pill for those who are opioid-naive, especially children. While documented reports of harm to patients by healthcare professional are few (most likely because professional healthcare workers are aware of those medications, such as cytotoxics and teratogens, that pose the greatest hazard if modified), the potential clearly exists. Some tablets are specially coated not to modify the absorption or release characteristics, but rather to prevent dermal contact or pulmonary exposure when handling. Breaking this coating poses an acute hazard to anyone in close proximity; residues can then disperse to the surrounding environment. Examples include cytotoxics (e.g., methotrexate, tamoxifen), steroids, prostaglandin analogs, and other hormones. An updated compilation of medications that should not be crushed, not just for therapeutic reasons, but also for safety concerns, is available at the Institute for Safe Medication Practices website ([106], http://www.ismp.org/tools/donotcrush.pdf.). An example of the cautions issued on this website include the one for finasteride (Propecia® and Proscar®.

This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).

Merck) and dutasteride (Avodart<sup>®</sup>, GlaxoSmithKline): "drug may cause fetal abnormalities; women who are, or may become, pregnant, should not handle capsules; all woman should use caution in handling capsules, especially leaking capsules." Oxymorphone (Opana ER<sup>®</sup>, Endo), oxycodone (OxyContin<sup>®</sup>, Purdue Pharma) and tramadol (Ultram ER<sup>®</sup>,Ortho-McNeil Pharmaceutical) users are warned, "tablet disruption can lead to rapid release and absorption of a potentially fatal dose of oxymorphone, oxycodone, or tramadol." The website offers advice about handeling Hydroxyurea (Droxia and Hydrea, Bristol-Myers Squibb) capsules: "exposure to powder may cause serious skin toxicities"; and lenalidomide (Revlimid, Celgene) capsules, calling it a "teratogenic analog of thalidomide."

While some of the dangers in the practice of physical drug destruction pertain solely to the administration of healthcare, some are also pertinent to the relatively recent recommended practice of crushing leftover, unwanted medications in order to facilitate their disposal, such as these instructions on the SmartRx website: ([102], http://www.smarxtdisposal.net): "Pour medication into a sealable plastic bag. If medication is a solid (pill, liquid capsule, etc.), crush it or add water to dissolve it." The intent of this recommendation is to render the medication useless to others so that disposal via trash does not lead to subsequent diversion by others. The consumer, lacking the knowledge of a healthcare professional, would probably not be aware of those medications that would pose acute risks from mechanical alteration – such as by crushing or opening capsules. The average consumer really has no way to know which pills are safe to mechanically destroy and which are dangerous – without carefully reading manufacturers' instructions. Many drugs could possibly be safely crushed (with the proper equipment), but since many should not be altered, comprehensive guidance for disposal would get complicated, as it has always been with respect to the ultimate route of disposal, where certain select medications (e.g.,

This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).

those with extreme toxicity or potential for abuse) should still be flushed into sewers to prevent unintended poisonings [23, 107]). Any additional handling of medications, beyond what is needed for therapeutic use, poses added risks for those in proximity and for the environment.

Crushing tablets or opening capsules by the consumer should also be discouraged for a variety of other reasons in addition to the immediate hazard to the person disposing of the medication in this manner. Some of these medications are formulated expressly to resist crushing. Mechanical destruction of medications can be time consuming and difficult. Some tablets can be extremely difficult to crush because of coatings or other properties designed purposefully to prevent alteration (making them physically impenetrable), and capsules can resist disassembly. Frustration could cause the consumer to rush the process or use excessive force, and as a result, disperse or spill dust, particles, or entire pills into the air, on the floor, countertop, or other surfaces or containers that might come into contact with food or beverages; sudden crushing of capsules could expel liquid contents. The dispersed API (or misplaced pills) could then come in contact with pets, infants, toddlers, and other unsuspecting people, where dermal, oral, and pulmonary exposures could occur.

Few consumers even have a mortar and pestle, much less a device specially designed to crush pills, such as those sometimes used in healthcare. They therefore will resort to any number of other improvised approaches, all of which require manual strength and dexterity, and which greatly increase the chances of local area contamination via spillage or dispersal: hammers, knives, pill splitters, spoon bottoms, nested spoons, rolling pins, etc. If a dedicated crushing device is used, it might also serve double duty for subsequent food preparation (e.g., a mortar and pestle used for spices) or eating (e.g., a spoon or cutting board). Yet another exposure pathway is then created. Consumers are often creative, and might also resort to other methods such as food

blenders; we have even handled an inquiry from a consumer who had planned to bake their medications in an oven, a practice that could result in pulmonary exposure to highly toxic chemicals. Once the hazards are understood, for consumers still wishing to crush pills, perhaps the best economical approach would be to use a heavy-duty commercial device specially designed for the purpose of crushing ([108], http://www.abinn.com) and where the crushed medications are collected directly into a plastic bag, thereby preventing accidental dispersal. For healthcare workers who want to dispose of larger quantities by crushing, electric automated crushers are available with automatic containment.

Furthermore, encouraging the consumer to mechanically destroy or to make unpalatable medications disposed via trash, "Mix drugs with an undesirable substance, such as cat litter or used coffee grounds...." ([23], http://www.whitehousedrugpolicy.gov/publications/pdf/prescrip\_disposal.pdf), may only provide an illusion of preventing reuse and diversion. Addicts and those who abuse drugs are known to be extremely persistent and clever at reclaiming drugs from all sorts of dirty matrices [105]. Any additional step or manipulation recommended for disposal of leftover medications incurs additional risk that medication can fall unnoticed onto floors or counters. Mixing with other substances (such as cat litter) prior to trash disposal is also not without controversy in the pharmacy community [109]; mixing with used cat litter also poses a risk of exposure to pathogen-laden dust.

The continuing need to flush or destroy those medications (both new and partially used) that are prone to diversion and abuse could possibly be avoided with advancements in formulation technologies for these drugs. There are a number of approaches under development; several, for example, are specifically designed to deter abuse of oral-use opiates [110]. Some examples

926

927

928

929

930

931

932

933

934

935

936

937

938

939

940

941

942

943

944

945

946

947

948

include: formulations that resist crushing and dissolving to obtain an injectable form, where the API is released very slowly and dissolving does not yield an injectable form (the oxycodone formulation Remoxy<sup>®</sup>. Pain Therapeutics) is an example: formulations having additives that cause unpleasant side effects if taken orally at supratherapeutic doses or if administered by a nontherapeutic route such as by injection or nasally (an example is the oxycodone formulation Acurox<sup>®</sup>, Acura Pharmaceuticals); and an approach that uses an opioid antagonist (such as naltrexone) in an indigestible form that cannot be absorbed if taken orally as designed, but which is readily released if the medication is crushed. An example is the Embeda® (Alpharma) formulation of morphine. The literature often mentions the use in Britain of drug destruction kits, usually referred to as DOOP kits, sometimes called controlled drug destruction (or denaturing) kits. While DOOP stands for Destruction Of Old Pharmaceuticals, the process employed has nothing to do with actual destruction or denaturing of the API chemical structure, but rather refers to the physical form of the medication. The process involves physically destroying the medication (e.g., crushing pills or emptying capsules) and mixing with a liquid that solidifies and serves to merely encapsulate the APIs. The point of emphasis here is that this approach would not, as its name implies provide a means for consumers to destroy APIs. The chemical destruction of APIs has been investigated as an alternative approach to incineration and for dealing with small quantities of waste drugs, especially the highly toxic antineoplastics ([111], http://www.who.int/injection\_safety/toolbox/docs/en/waste\_management.pdf; [112], http://www.noharm.org/library/docs/NoIncineration Medical Waste Treatment Techn.pdf). These approaches have generally involved the use of concentrated acids and oxidants, such as

949

950

951

952

953

954

955

956

957

958

959

960

961

962

963

964

965

966

967

968

969

970

971

permanganate, sodium hypochlorite, hydrogen peroxide (also with iron - Fenton's reagent), sulfuric acid, nitric acid, and hydrochloric acid. They also usually involve heating and are quite hazardous. Different methods seem to be required for different APIs. One universal approach for all APIs has never been proposed. Often proposed as a means of on-site destruction of APIs at drug collection events (with the intent of permitting the return of controlled substances), chemical destruction is not yet feasible for widespread implementation because of: the hazardous nature of the procedures, the fact that the complete destruction of all APIs cannot be assured (a Drug Enforcement Administration requirement for controlled substances), and the unknowns with regard to the possible generation of hazardous by-products (especially those that are volatile) as a result of multiple APIs undergoing many reactions simultaneously. Greener, lesshazardous destruction methods are just beginning to be developed. These use comparatively lesshazardous reagents and generate much less hazardous waste. One example is the use of an irontetraamidomacrocyclic ligand (Fe-TAML) in conjunction with hydrogen peroxide, which has proved highly effective at destroying a variety of APIs, such as estrogens [113]. Another might be the use of electrolysis [114].

Certain drugs should not be unnecessarily handled or altered by consumers, especially those that are considered hazardous. In an occupational setting, hazardous drugs should be handled only when proper containment of dusts, particles, and vapors is sufficient. Crushing tablets or opening capsules containing hazardous drugs should be avoided, even by compounding pharmacists and other healthcare professionals; this points to the heightened hazards that pill alteration could pose to untrained and ill-equipped consumers. Overviews of hazardous drugs and guidelines for their proper and safe handling are available ([115],

http://www.cdc.gov/niosh/docs/2004-165/pdfs/2004-165.pdf; [116, 117]). A subset of these pose

972

973

974

975

976

977

978

979

980

981

982

983

984

985

986

987

988

989

990

991

992

993

994

risks with respect to dermal or pulmonary exposure (via particulates, dusts, or powders), although for healthcare workers, dermal contact with these drugs (from patients or other sources) is a primary route of exposure, possibly via subsequent hand-to-mouth contact [117].

In the U.S., unintentional poisoning by medications is a leading cause of injury in children (ages 18-35 months). The types of medications commonly involved with poisonings are summarized by Meyer et al. [118]. These are among the medications for which mechanical alteration would pose the highest risk as a result of inadvertent dispersal of particles or whole doses. Those drugs commonly involved with non-life threatening poisonings include: antibiotics. B2 agonists and sympathomimetics (e.g., phenylephrine and ephedrine), and non-steroidal antiinflammatories (e.g., mefenamic acid and phenylbutazone). Those involved with a high potential for adverse effects include: antihistamines (H1 and H2 receptor antagonists), beta-blockers, calcium channel blockers (e.g., dihydropyridines such as nifedipine), phenylalkylamines such as verapamil, benzothiazepines (e.g., diltiazem), digoxin, isoniazid, sulfonylureas, and tricyclic antidepressants. Those with the potential for life-threatening effects from even small doses (such as the equivalent of a single, non-delayed release tablet) include: calcium channel blockers, chloroquine/hydroxychloroquine, clonidine, clozapine/olanzapine, flecainide, imidazolines, loxapine, opioids, phenothiazines (thioridazine and chlorpromazine), quinine, sulfonylureas, theophylline, and tricyclic antidepressants (amitryptyline, imipramine and desipramine); also see Bar-Oz et al. [119]. Still others are noted for delayed effects that might not be immediately noticed: diphenoxylate and atropine, hypoglycemic agents, monoamine oxidase inhibitors, and acetaminophen (larger quantities).

Some caveats are also important regarding the guidance issued by manufacturers and others [23, 107] on the disposal of certain hazardous medications by the consumer. The manufacturer's

instructions themselves can cause confusion. One example is the disposal of fentanyl formulated in transmucosal delivery systems such as oral (buccal) lozenges or handles (lollipops). The residue that remains on the handle itself (which should not be flushed) or in partially used lozenges varies greatly, but can be hazardous. Disposal instructions call for dissolving the residue of partially or completely used doses by holding under running hot water. An opiate-naive person could possibly absorb a toxicologically significant dose of fentanyl through the skin (especially if open wounds were present) if partially used lozenges or handles were held with exposed fingers during this process.

A final concern regarding destruction prior to disposal via trash is if someone were poisoned by accidental or purposeful ingestion of crushed pills reclaimed from the trash or from spillage, it would not be as simple and fast to identify the responsible medication as it would be if the intact medication (with identifying information) were available.

#### Disposal

The wastage caused by unused, leftover medications was recognized as early as the 1970s with some evaluations of the types and quantities of medications returned by the public [120, 121]. The processes developed over the last three decades for handling drug waste generated by consumers has varied greatly among countries [3, 10, 11]. In the U.S., despite federal guidelines ([23], http://www.whitehousedrugpolicy.gov/publications/pdf/prescrip\_disposal.pdf), confusion and debate surround what constitutes the best approaches for disposal ([24, 122], http://blog.epa.gov/blog/2008/12/08/qotw-prescription-drug-disposal).

The overall significance of disposal of medications with respect to its contribution of individual APIs (and APIs in general) is an unresolved question [15]. The information needed to

This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).

make this type of assessment has not been available. Here we refer to the relative significance of disposal (versus excretion) as the relative environmental footprint (REF) of a disposed drug. As used here, the REF does not refer to the impact in the environment of the API, but rather to the potential importance of disposal. The REF for disposal is a simplified form of the more comprehensive relative significance (Sr) factor discussed earlier.

Similar to Sr, a disposed drug's relative environmental footprint (REF<sub>d</sub>) is a function of two factors: the fraction of overall API mass (or moles) disposed via sewers, and the fraction of API that is excreted unchanged (and/or washed from the skin). The REF<sub>d</sub> for a particular API is defined as the contribution of an API to sewage by disposal relative to that released from intended usage (such as via excretion). This can be calculated on the basis of either mass or moles of API as:

# [fraction disposed]/[fraction excreted] = $REF_d$ (3)

Here are some hypothetical examples. Assuming that 10% of an API is excreted and 5% is disposed, then disposal of 1 dose would be equivalent to consuming 0.5 doses with respect to the introduction of the API to sewage. Similarly, assuming that 80% of an API is excreted and 5% is disposed, then disposal of 1 dose would be equivalent to consuming 0.06 doses. With 1% excreted and 5% disposed, disposal would be equivalent to consuming 5 doses. And with 0.01% excreted and 95% disposed (e.g., the remaining residue retained in the container/dispenser), disposal would be equivalent to consuming 9,500 doses. The latter example might emulate the case for an API that was used almost exclusively in topical preparations (and with nominal systemic absorption), which would then be washed from the skin during bathing. So the possible range for REF<sub>d</sub> values can range from near zero (where disposal is a non-factor in contributing an API to the environment) to extremely high (where disposal is a major factor).

While  $REF_d$  provides the relative potential contributions by disposal among APIs, in order to gain an understanding of the actual magnitude of API release via disposal, the  $REF_d$  must be multiplied by the actual number of doses sold or dispensed (during a defined period of time):

When multiplied by the number of doses (ND), the  $REF_d$  for a particular API yields the hypothetical number of consumed doses that would be required to release the equivalent amount of API actually contributed by disposal:

 $REF_d \bullet ND/time = equivalent doses contributed by disposal during a defined time period (4)$ 

The number of doses must be calculated on the basis of the same units as  $REF_d$  (either mass or moles). Note that when ranking drugs according to  $REF_d$  equivalent doses, the relative ranking could change depending on whether the  $REF_d$  is calculated on the basis of mass or moles; low-molecular-weight APIs would yield larger numbers of doses when expressed in terms of moles, and high-molecular-weight APIs yielding more doses when the  $REF_d$  is expressed in terms of mass.

A ranking of REF<sub>d</sub> values for various APIs would not necessarily be in the same order as the ranked actual contributions. For example, disposal of a high REF<sub>d</sub> API for an infrequently prescribed medication might contribute less API than a medication whose REF<sub>d</sub> is comparatively very low but which is frequently prescribed.

The variance in the REF<sub>d</sub> for a given API will be most affected by the rate of disposal, which could vary wildly as a function of many variables [8]. The rate of disposal might be most affected by the type of drug or its therapeutic class, some of which have much greater rates of non-compliance than others. The REF<sub>d</sub> could allow inter-comparisons of drugs to determine the relative importance of disposal with respect to contributing to the occurrence of their respective APIs in sewage. By converting numbers of doses to daily doses in terms of mass or moles, a

This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).

direct relationship with potency can be obtained. If human potency were assumed to correlate with the potential for ecological effects, then REF<sub>d</sub> could be used to reveal which APIs are being disposed in amounts having the highest potential for ecological effects.

The REF<sub>d</sub> can be understood best by considering some extreme examples. A drug that is disposed in relatively large quantities can nonetheless have a comparatively low REF<sub>d</sub> if its overall use is comparatively larger and/or if it is excreted largely unchanged (extensively excreted). A drug that is disposed in relatively small quantities can have a comparatively higher REF<sub>d</sub> if its overall use is comparatively smaller and/or if it is extensively metabolized (leaving little to be excreted unchanged). For an extensively excreted API, both disposal and excretion contribute equally to the environmental loading of the API (each pill disposed contributes to the environmental load the same as if the pill were ingested). For externally applied APIs that are poorly absorbed, the significance of disposal is a direct function of the portion disposed versus the portion absorbed after its designed use (each dose disposed contributes to the environmental load the same as if the dose were applied externally as intended but not absorbed).

The two extreme scenarios that maximize and minimize the significance of disposal are, respectively: disposal of a large fraction of an API that would otherwise be extensively metabolized, and disposal of a small fraction of a drug that would otherwise be excreted largely unchanged (or of topical drugs that are poorly absorbed). The former is exacerbated when the API is purchased in large quantities, and the latter is attenuated yet further when the API is purchased in small quantities.

Five generalizations can be made. Disposal of APIs that would otherwise be extensively metabolized will tend to be responsible for larger percentages of the API in the environment. Disposal of APIs that would otherwise be extensively excreted unchanged will tend be

responsible for smaller percentages of the API in the environment. Models that use the default assumption of extensive excretion (no metabolic conversion) for APIs that actually undergo extensive metabolism will greatly underestimate contributions from disposal. For APIs applied dermally or by delivery devices, the significance of disposal is a direct function of the portion disposed versus the portion absorbed after intended usage. The REF<sub>d</sub> is maximized when 100% is disposed and/or 100% is metabolized (none excreted unchanged).

Three major questions could be addressed with this approach. For those APIs that are most frequently detected by environmental monitoring (and in the highest concentrations), do they also have high REF<sub>d</sub> values? These might have higher contributions from disposal. For those APIs that are monitored for but rarely detected, are they also among the ones with the lower REF<sub>d</sub> values and that are extensively metabolized? These might have little contribution from disposal. Are there APIs with high REF<sub>d</sub>'s that have never been monitored for? If so, these might be likely targets for monitoring. If detected at critical concentrations, these are also the drugs that might be likely targets for stringent controls on disposal.

Dermal excretion, dermal application, disposal, and lack of absorption from the gut may well explain the presence in sewage of those APIs that are otherwise extensively metabolized; note, however, that this does not take into consideration the extensive conjugation that many drugs undergo, which can be followed by bacterial deconjugation to return the parent drug [59].

In fact, alternative sources of API pollutants that have attracted little attention (such as dermal application, excretion via sweat, and disposal to sewers) may have already served to confuse the conclusions reached by some regarding the presence of certain APIs in the environment. For example, Jjemba [123] reported a possible negative correlation (based on a very small data set) between the efficiency of excretion of an API in its unmetabolized, parent form

and its occurrence in the environment: "the drugs that have a low proportion of the parent

1110

1111

1112

1113

1114

1115

1116

1117

1118

1119

1120

1121

1122

1123

1124

1125

1126

1127

1128

1129

1130

1131

1132

compound excreted also display a higher concentration in the aquatic environment". Noted was the widespread occurrence of poorly excreted APIs (e.g., acetylsalicylic acid [aspirin], ibuprofen, acetaminophen [paracetamol], and carbamazepine), as well as some moderately excreted APIs (e.g., sulfamethoxazole, diclofenac, primidone, and rinitinide [sic, ranitidine]). All but three of these, however, are available OTC and are purchased in large quantities, which makes them prone to expiration and subsequent disposal. Ibuprofen, acetaminophen, and diclofenac have been reported as among the unused drugs most frequently returned to pharmacies by consumers [124]. Since drugs returned to take-back events currently represent such a small percentage of those that are otherwise disposed to sewers and trash, this poses the possibility that perhaps even larger quantities of these drugs are disposed to sewers. Indeed, in Ruhoy's study of medication disposal inventories assembled from a coroner's office ([125], http://environment.unlv.edu/abstractsGrad/ruhoy.html) acetaminophen and ibuprofen were the first and ninth most abundant medications disposed over a 1-year period (S. Ruhoy, unpublished data). Both are extensively conjugated or oxidized with little unchanged API excreted. The negative correlation noted by Jiemba [123] might simply be the result of not considering all of the possible sources. Another example is the fifth-most abundant API disposed in the inventory of coroner data conducted by Ruhoy (unpublished data), carisoprodol. Extensive metabolism yields at least three active metabolites (one of which is meprobamate); only traces of carisoprodol appear in the urine [126]. Despite being extensively metabolized, carisoprodol has been reported in several recent monitoring studies. It was even reported at 129 ng/L in recycled water [127]. It was the only API identified in recycled water (at up to 217 ng/L), and also reported in secondary effluent

1133 ([128],

http://www.valleywater.org/website/media/pdf/Streamflow%20AugmentationDraft%20IS%20M ND.pdf ). It has also been tentatively identified in runoff from fields irrigated with treated wastewater or effluent-dominated stream water [129]. Note, however, that some conditions can cause the excretion of unchanged carisoprodol (as with many other APIs); these include concurrent administration of APIs that inhibit microsomal oxidases, certain polymorphisms in microsomal oxidases, and stress, which reduces absorption from the gut. Nonetheless, carisoprodol is an example of an API for which disposal might be playing a dominant role in its environmental occurrence.

# Disposal of problematic medications

The continuing need in the U.S. to dispose of certain medications by flushing to sewers and actively avoiding disposal in the trash [23, 107], at least until sustainable take-back or collection programs are developed, poses a dilemma in balancing the protection of human health and safety with protection of the environment [16]. These are the highly hazardous medications or those subject to abuse that could be diverted or accidentally acquired if they were disposed in the trash. The potential significance of disposal by flushing can be evaluated by examining the pharmacokinetics of this subgroup of medications to determine the fraction of the API that is excreted unchanged. But two other factors are also required to determine the relative significance of disposal among APIs: the total amount of drug purchased and the fraction eventually disposed (Table 6).

In the absence of data for these two factors, a preliminary idea can be formulated as to which drugs that currently require flushing are contributing the highest percentage of APIs to the

environment. An expanded list of drugs (but still not comprehensive) where disposal via flushing is recommended [107] is shown in **Table 7**. These are annotated with information regarding whether they are extensively metabolized or excreted unchanged.

If the efficiency with which an API is excreted unchanged is high, then contributions by disposal would have comparatively less impact (unless an inordinate percentage of the drug is disposed versus actually being used; for example if a medication experiences inordinate non-compliance among patients). On the other hand, if the excretion efficiency is very low (extensive metabolism), then disposal might play an important contributory role. These would be the drugs that could be targeted for alternative approaches for safe disposal should their APIs prove hazardous for the environment.

The sub-group of drugs (those where sewer disposal is still recommended) for which disposal has the potential to play a dominant role in contributing to environmental residues is annotated in Table 7. Note, however, that this assessment ignores the possible contributions from hydrolyzable conjugates or bioactive metabolites excreted in urine/feces. From this assessment, the limited sub-group for which disposal might play only a minor role as a source of APIs in the environment include: entecavir (Baraclude®, Bristol Myers Squibb); oxymorphone (Opana®/Opana ER®, Endo Pharmaceuticals); buprenorphine (Suboxone® & Subutex®, Reckitt Benckiser); gatifloxacin (Tequin®, Bristol Myers Squibb); telbivudine (Tyzeka®, Novartis Pharma Stein AG) and stavudine (Zerit®, Bristol-Myers Squibb).

These are the medications whose continued flushing would most likely contribute the *least* to environmental loadings for the APIs they contain. Until suitable disposal alternatives are available, consideration could be given to continue advising the flushing of these drugs in order to ensure they do not contribute to poisonings or abuse. Human safety concerns (from unsecured

This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).

disposal, such as in trash) might clearly outweigh the probably negligible risks for the environment (from disposal to sewers).

1179

1180

1181

1182

1183

1184

1185

1186

1187

1188

1189

1190

1191

1192

1193

1194

1195

1196

1197

1198

1199

1200

1201

This approach, however, only conveys the potential role that disposal could play. Two other factors required in determining the significance of disposal are the fraction of total drug purchased that is eventually disposed and the overall usage rate of the medication (Table 6). Unfortunately, disposal data are very rare. Disposal data were collated from the records collected and maintained by the Clark County Coroner (Las Vegas, NV, USA) using the approach described by Ruhoy and Daughton [3]. These data are located in last column of Table 7. By cross checking the two sets of data (i.e., the potential for disposal significance, based on PK data, versus actual disposal data), the following drugs are the ones that are disposed in the largest quantities (in Clark County Nevada over a particular one-year period) and which also have the potential for contributing the larger portions of APIs to the environment: morphine sulfate extended-release (Avinza<sup>®</sup>, Ligand Pharmaceuticals); meperidine (Demerol®, Sanofi-Synthelabo); methadone (Dolophine®, Roxane Laboratories); oxycodone (OxyContin<sup>®</sup>, Purdue Pharma and Percocet<sup>®</sup>, Endo Pharmaceuticals); and atazanavir (Revataz<sup>®</sup>, Bristol-Myers Squibb); morphine is excreted largely as conjugates, and therefore disposal's contribution might not be appreciable. These are the medications for which alternative disposal practices (other than flushing) might be beneficial for the environment. Note that in 2008, the manufacturer re-labeled Reyataz and Baraclude and no longer recommends flushing [130].

Note that fentanyl, although not in the coroner's inventory in substantial quantities, would also be among the drugs where disposal to sewage could prove to be an important source, since it is extensively metabolized. Also note that for drugs administered via a delivery device (e.g., patches, lollipops) or dermally, there will always be additional wastage (residue remaining in the

device). These residues, which can be substantial for patches or partially used lozenges, essentially serve as another contributor analogous to disposal.

For those drugs in Table 7 that were not recovered in the coroner inventory (buprenorphine, diazepam, didanosine, entecavir, methylphenidate, oxymorphone, sodium oxybate, telbivudine), this could indicate that these drugs are sold in very small quantities or that patient compliance tends to be very high – either scenario not promoting leftovers. For these drugs, guidance to dispose by flushing may be inconsequential with respect to environmental impact resulting from excreted residues. It is important to emphasize that these assessments are based on only one study of actual disposal practice and need to be corroborated by further studies (e.g., using data collected from coroner offices in other locales).

#### Conclusion

It has been long assumed that the active ingredients from human pharmaceuticals (APIs) enter the environment as trace pollutants primarily as a result of their excretion via urine and feces. Urine conveys portions of the APIs that escape metabolism as well as conjugates that are susceptible to later hydrolysis (returning the parent form of the API) and other metabolites (some of which can be highly bioactive). The feces convey metabolites excreted via the bile as well as those portions of APIs that are not absorbed from oral medications.

For the first time, several alternative routes for the entry into the environment by way of sewage have been shown to possibly be important for certain APIs (or therapeutic classes) having particular pharmacokinetic parameters or usage characteristics. These routes include: release of APIs from skin during bathing and washing (those applied topically or transdermally as well as those excreted to the skin via sweat), disposal of unused, leftover medications, and disposal of

This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).

used and partially used medical devices, especially transdermal delivery systems. The published literature relevant to these alternative routes has been compiled for the first time, and examples of drugs for which these routes are possibly important are presented.

Routes other than drinking water and foods by which humans can be directly and inappropriately exposed to chronic and acute doses of APIs are also discussed. These include: direct interpersonal dermal transfer; indirect exposure via contact with items touched or used by those who are medicated (e.g., door knobs, telephones, clothing, spas), accidental exposure (such as ingestion by infants, toddlers, or pets) and inappropriate reuse (or abuse) of used or partially used transdermal devices, and unintended exposure to dust, particulates, and scattered pills/capsules that consumers unwisely attempt to destroy (such as by crushing, a recommendation made by some organizations wanting to keep abused drugs from being diverted) before disposing in trash. Some of these routes are documented as leading to morbidity or mortality. All of these routes are interconnected in the lifecycle of APIs in the environment (see Fig. 1).

The unifying concept of pharmEcokinetics was introduced as the umbrella under which the interrelationships can be understood. Some of the vulnerabilities in the lifecycle of an API present opportunities for pollution prevention, an example being more efficient and better targeted delivery of transdermal APIs; proper education of patients by prescribers and pharmacists regarding the application of topical products might also help to reduce over usage. Current recommendations regarding the disposal of certain highly abused drugs by flushing into sewers may not contribute substantially to APIs in sewers; until disposal alternatives are made available more prudent than domestic trash, a rationale is presented for the continuation of disposing these particular medications to sewers. Predictive models that assume extensive excretion for APIs that actually undergo extensive metabolism will greatly underestimate the significance of disposal.

This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).

1249

1250

1251

1252

1253

1254

1255

1256

1257

1258

1259

1260

1261

1262

1263

1264

There are many variables that determine the overall significance of these secondary transport and exposure routes. While none of these routes has been realistically factored into published exposure, transport, or fate models (other than in a general way, using generic assumptions), the present study should facilitate the collection of the needed data to make models more accurate and useful, especially for basing decisions involved with pollution prevention or source control. Acknowledgment-Ilene Ruhoy thanks David Hassenzahl, University of Nevada, Las Vegas for continued guidance, and the U.S. Environmental Protection Agency for granting an appointment as a U.S. EPA Student Volunteer. We also thank Sheila King (Senior Environmental Employment Program, United States Environmental Protection Agency, Las Vegas) for her diligent maintenance of the EndNote PPCPs Literature Database. The authors greatly appreciate the efforts of the three anonymous reviewers, who invested considerable time and thought in providing extensive comments that greatly improved the quality of the final product. The U.S. Environmental Protection Agency through its Office of Research and Development funded and managed the research described here. It has been subjected to the Agency's administrative review and approved for publication as an U.S. EPA document.

### REFERENCES

1265

- 1266 [1] U.S. Environmental Protection Agency. 2008. Pharmaceuticals and Personal Care
- Products (PPCPs): Relevant Literature. Office of Research and Development, Las Vegas, NV.
- 1268 [2] Kolpin DW, Furlong ET, Meyer MT, Thurman EM, Zaugg SD, Barber LB, Buxton HT.
- 1269 2002. Pharmaceuticals, hormones, and other organic wastewater contaminants in U.S. streams,
- 1270 1999-2000: A national reconnaissance. Environ Sci Technol 36:1202-1211.
- 1271 [3] Daughton CG. 2007. Pharmaceuticals in the environment: Sources and their management.
- 1272 In Petrovic M, Barcelo D, eds, Analysis, Fate and Removal of Pharmaceuticals in the Water
- 1273 Cycle, Vol 50–Wilson & Wilson's Comprehensive Analytical Chemistry Series. Elsevier
- 1274 Science, Amsterdam, The Netherlands, pp 1-58.
- 1275 [4] Winker M, Faika D, Gulyas H, Otterpohl R. 2008. A comparison of human pharmaceutical
- 1276 concentrations in raw municipal wastewater and yellowwater. Sci Total Environ 399:96-104.
- 1277 [5] Boxall ABA. 2002. The environmental side effects of medication How are human and
- veterinary medicines in soils and water bodies affecting human and environmental health?
- 1279 *EMBO Reports* 5:1110-1116.
- 1280 [6] Boxall ABA, Fogg LA, Blackwell PA, Kay P, Pemberton EJ, Croxford A. 2004.
- 1281 Veterinary medicines in the environment. Rev Environ Contam Toxicol 180:1-91.
- 1282 [7] Cunningham VL, Buzby M, Hutchinson T, Mastrocco F, Parke N, Roden N. 2006. Effects
- of human pharmaceuticals on aquatic life: Next steps. *Environ Sci Technol* 40:3456-3462.
- Ruhoy I, Daughton C. 2008. Beyond the medicine cabinet: An analysis of where and why
- medications accumulate. *Environ Int*34:1157-1169.

- This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).
- 1286 [9] Daughton CG. 2003. Cradle-to-cradle stewardship of drugs for minimizing their
- environmental disposition while promoting human health. I. Rationale for and avenues toward a
- green pharmacy. Environ Health Perspect 111:757-774.
- 1289 [10] Daughton CG. 2003. Cradle-to-cradle stewardship of drugs for minimizing their
- environmental disposition while promoting human health. II. Drug disposal, waste reduction, and
- future directions. *Environ Health Perspect* 111:775-785.
- 1292 [11] Glassmeyer S, Hinchey E, Boehme S, Daughton C, Ruhoy I, Conerly O, Daniels R, Lauer
- L, McCarthy M, Nettesheim T, Sykes K, Thompson V. 2009. Disposal Practices for unwanted
- residential medications in the United States. *Environ Int* 35:566-572.
- 1295 [12] Bound JP, Kitsou K, Voulvoulis N. 2006. Household disposal of pharmaceuticals and
- perception of risk to the environment. *Environ Toxicol Pharmacol* 21:301-307.
- 1297 [13] Doerr-MacEwen NA, Haight ME. 2006. Expert stakeholders' views on the management of
- human pharmaceuticals in the environment. *Environ Manag* 38:853–866.
- 1299 [14] Seehusen DA, Edwards J. 2006. Patient practices and beliefs concerning disposal of
- medications. Journal of the American Board of Family Medicine 19:542-547.
- Ruhoy IS, Daughton CG. 2007. Types and quantities of leftover drugs entering the
- environment via disposal to sewage Revealed by coroner records. Sci Total Environ 388:137-
- 1303 148.
- 1304 [16] Daughton CG, Ruhoy IS. 2008. The afterlife of drugs and the role of PharmEcovigilance.
- 1305 Drug Saf 31:1069-1082.
- 1306 [17] Fasola G, Aita M, Marini L, Follador A, Tosolini M, Mattioni L, Mansutti M, Piga A,
- Brusaferro S, Aprile G. 2008. Drug waste minimisation and cost-containment in Medical
- Oncology: Two-year results of a feasibility study. BMC Health Serv Res 8:70.

- 1309 [18] National Cancer Institute. 2008. Dictionary of Cancer Terms: Pharmacokinetics. National
- 1310 Institutes of Health, Washington, DC, USA.
- 1311 [19] Castegnaro M, Hansel S. 2006. Antineoplastic drugs in sewage from hospitals and
- households. Environment, Risques & Santé 5:266-270.
- 1313 [20] Inciardi JA, Surratt HL, Kurtz SP, Cicero TJ. 2007. Mechanisms of prescription drug
- diversion among drug-involved club- and street-based populations. *Pain Med* 8:171-183.
- 1315 [21] Burt A, Annest J, Ballesteros M, Budnitz D. 2006. Nonfatal, unintentional medication
- exposures among young children--United States, 2001-2003. JAMA 295:882-884.
- 1317 [22] Wysowski DK. 2007. Surveillance of prescription drug-related mortality using death
- certificate data. *Drug Saf* 30:533-540.
- 1319 [23] Office of National Drug Control Policy. 2007. Proper disposal of prescription drugs.
- 1320 Washington, DC, USA.
- 1321 [24] Code of Federal Regulations, Drug Enforcement Administration. 2008. Disposal of controlled
- substances by persons not registered with the Drug Enforcement Administration. 21 CFR Parts 1300,
- 1323 1301, 1304, 1305, and 1307, 74 (12):3480-3487. U. S. Government Printing Office, Washington,
- 1324 DC.
- 1325 [25] Kostich MS, Lazorchak JM. 2008. Risks to aquatic organisms posed by human
- pharmaceutical use. *Sci Total Environ* 389:329-339.
- 1327 [26] Cooper ER, Siewicki TC, Phillips K. 2008. Preliminary risk assessment database and risk
- ranking of pharmaceuticals in the environment. Sci Total Environ 398:26-33.
- 1329 [27] Anderson PD, D'Aco VJ, Shanahan P, Chapra SC, Buzby ME, Cunningham VL,
- DuPlessie BM, Hayes EP, Mastrocco FJ, Parke NJ, Rader JC, Samuelian JH, Schwab BW. 2004.

- This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).
- Screening analysis of human pharmaceutical compounds in U.S. surface waters. *Environ Sci*
- 1332 Technol 38:838-849.
- 1333 [28] Wilkosz MF, Bogner RH. 2003. Transdermal drug delivery-Part 1: Current status. US
- 1334 *Pharm* 28:40-55.
- 1335 [29] Fisher PMJ, Scott R. 2008. Evaluating and controlling pharmaceutical emissions from
- dairy farms: A critical first step in developing a preventative management approach. J Clean
- 1337 *Prod* 16:1437-1446.
- 1338 [30] Thaysen JH, Schwartz IL. 1953. The permeability of human sweat glands to a series of
- sulfonamide compounds. *J Exp Med* 98:261-268.
- 1340 [31] Rouen D, Dolan K, Kimber J. 2001. A review of drug detection testing and an examination
- of urine, hair, saliva and sweat. Technical Report 120. National Drug and Alcohol Research
- 1342 Centre, University of New South Wales, Sydney, Australia.
- 1343 [32] Fortner NA. 2008. The detection of drugs in sweat. In Jenkins AJ, ed, Forensic Science
- and Medicine: Drug Testing in Alternate Biological Specimens. Humana Press, Totowa, NJ,
- 1345 USA, pp 101-116.
- 1346 [33] Parnas J, Flachs H, Gram L, Wurtz-Jorgensen A. 1978. Excretion of antiepileptic drugs in
- 1347 sweat. *Acta Neurol Scand* 58:197-204.
- 1348 [34] Leggett R, Lee-Smith EE, Jickells SM, Russell DA. 2007. "Intelligent" Fingerprinting:
- simultaneous identification of drug metabolites and individuals by using antibody-functionalized
- nanoparticles. Angewandte Chemie International Edition 46:4100-4103.
- 1351 [35] Pichini S, Navarro M, Pacifici R, Zuccaro P, Ortuno J, Farre M, Roset PN, Segura J, de la
- Torre R. 2003. Usefulness of sweat testing for the detection of MDMA after a single-dose
- administration. J Anal Toxicol 27:294-303.

- 1354 [36] Tobin DJ. 2006. Biochemistry of human skin-our brain on the outside *Chem Soc Rev*
- 1355 35:52-67.
- 1356 [37] Preston KL, Huestis MA, Wong CJ, Umbricht A, Goldberger BA, Cone EJ. 1999.
- Monitoring cocaine use in substance-abuse-treatment patients by sweat and urine testing. J Anal
- 1358 *Toxicol* 23:313-322.
- 1359 [38] Joss A, Zabczynski S, Gobel A, Hoffmann B, Loffler D, McArdell CS, Ternes TA,
- 1360 Thomsen A, Siegrist H. 2006. Biological degradation of pharmaceuticals in municipal
- wastewater treatment: Proposing a classification scheme. Water Res 40:1686-1696.
- 1362 [39] Appenzeller BMR, Schummer C, Rodrigues SB, Wennig R. 2007. Determination of the
- volume of sweat accumulated in a sweat-patch using sodium and potassium as internal reference.
- 1364 *J Chromatogr B* 852:333-337.
- 1365 [40] Høiby N, Jarløv JO, Kemp M, Tvede M, Bangsborg JM, Kjerulf A, Pers C, Hansen H.
- 1366 1997. Excretion of ciprofloxacin in sweat and multiresistant Staphylococcus epidermidis. Lancet
- 1367 349:167-169.
- 1368 [41] Schneider S, Ait-m-bark Z, Schummer C, Lemmer P, Yegles M, Appenzeller B, Wennig
- 1369 R. 2008. Case report: Determination of fentanyl in sweat and hair of a patient using transdermal
- 1370 patches. *J Anal Toxicol* 32:260-264.
- 1371 [42] Daughton CG. 2008. Pharmaceuticals as environmental pollutants: The ramifications for
- human exposure. In Heggenhougen K, Quah S, eds, *International Encyclopedia of Public*
- 1373 *Health*, Vol 5. Academic Press, Oxford, UK, pp 66-102.
- 1374 [43] Martin ANF, George R, Jones AD, Frank M. 2007. The non-destructive identification of
- solid over-the-counter medications using single particle aerosol mass spectrometry. *Rapid*
- 1376 *Commun Mass Spectrom* 21:3561-3568.

- This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).
- 1377 [44] Takats Z, Wiseman JM, Gologan B, Cooks RG. 2004. Mass spectrometry sampling under
- ambient conditions with desorption electrospray ionization. *Science* 306:471-473.
- 1379 [45] Williams JP, Patel VJ, Holland R, Scrivens JH. 2006. The use of recently described
- ionisation techniques for the rapid analysis of some common drugs and samples of biological
- origin. Rapid Commun Mass Spectrom 20:1447-1456.
- 1382 [46] Høiby N, Pers C, Johansen HK, Hansen H, The Copenhagen Study Group on Antibiotics
- in Sweat. 2000. Excretion of β-lactam antibiotics in sweat—A neglected mechanism for
- development of antibiotic resistance? *Antimicrob Agents Chemother* 44:2855-2857.
- 1385 [47] Jacobi U, Waibler E, Schulze P, Sehouli J, Oskay-Ozcelik G, Schmook T, Sterry W,
- Lademann J. 2005. Release of doxorubicin in sweat: first step to induce the palmar-plantar
- erythrodysesthesia syndrome? *Annals of Oncology* 16:1210-1211.
- 1388 [48] Lorusso D, Di Stefano A, Carone V, Fagotti A, Pisconti S, Scambia G. 2007. Pegylated
- liposomal doxorubicin-related palmar-plantar erythrodysesthesia ('hand-foot' syndrome). Annals
- 1390 of Oncology 18:1159-1164.
- 1391 [49] Madsen E, Larsen H. 1988. Excretion of mutagens in sweat from humans treated with anti-
- neoplastic drugs. Cancer Lett 40:199-202.
- 1393 [50] Mader R, Rizovski B, Steger G, Wachter A, Kotz R, Rainer H. 1996. Exposure of
- oncologic nurses to methotrexate in the treatment of osteosarcoma. Arch Environ Health 51:310-
- 1395 314.
- 1396 [51] Fransman W, Vermeulen R, Kromhout H. 2005. Dermal exposure to cyclophosphamide in
- hospitals during preparation, nursing and cleaning activities. *Int Arch Occup Environ Health*
- 1398 78:403-412.

- This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).
- 1399 [52] Meijster T, Fransman W, Veldhof R, Kromhout H. 2006. Exposure to antineoplastic drugs
- outside the hospital environment. Ann Occup Hyg 50:657-664.
- 1401 [53] Connor TH, Shults M, Fraser MP. 2000. Determination of the vaporization of solutions of
- mutagenic antineoplastic agents at 23 and 37°C using a desiccator technique. *Mutation*
- 1403 Research/Genetic Toxicology and Environmental Mutagenesis 470:85-92.
- 1404 [54] Kiffmeyer T, Kube C, Opiolka S, Schmidt K, Schöppe G, Sessink P. 2002. Vapour
- pressures, evaporation behaviour and airborne concentrations of hazardous drugs: Implications
- 1406 for occupational safety. *Pharm J* 268:331-337.
- 1407 [55] Fransman W, Huizer D, Tuerk J, Kromhout H. 2007. Inhalation and dermal exposure to
- eight antineoplastic drugs in an industrial laundry facility. *Int Arch Occup Environ Health*
- 1409 80:396-403.
- 1410 [56] Fransman W. 2006. Antineoplastic drugs occupational exposure and health risks. PhD
- thesis. Utrecht University, Nijkerk, The Netherlands.
- 1412 [57] Monthly Prescribing Reference. 2008. Monthly Prescribing Reference Database:
- Dermatological disorders. Haymarket Media, New York, NY, USA.
- 1414 [58] Niazi SK. 2004. Handbook of Pharmaceutical Manufacturing Formulations: Semisolid
- 1415 *Products*, Vol 4. CRC, Boca Raton, FL, USA, pp 1-263.
- 1416 [59] D'Ascenzo G, Di Corcia A, Gentili A, Mancini R, Mastropasqua R, Nazzari M, Samperi R.
- 1417 2003. Fate of natural estrogen conjugates in municipal sewage transport and treatment facilities.
- 1418 Sci Total Environ 302:199-209.
- 1419 [60] Arcand-Hoy LD, Nimrod AC, Benson WH. 1998. Endocrine-modulating substances in the
- environment: Estrogenic effects of pharmaceutical products. *Int J Toxicol* 17:139-158.

- 1421 [61] Walters KA, Brain KR, Green DM, James VJ, Watkinson AC, Sands RH. 1998.
- 1422 Comparison of the transdermal delivery of estradiol from two gel formulations. *Maturitas*
- 1423 29:189-195.
- 1424 [62] Al-Dujaili EAS. 2006. Development and validation of a simple and direct ELISA method
- for the determination of conjugated (glucuronide) and non-conjugated testosterone excretion in
- 1426 urine. Clinica Chimica Acta 364:172-179.
- 1427 [63] Timón Andrada R, Maynar Mariño M, Muñoz Marín D, Olcina Camacho G, Caballero M,
- Maynar Mariño J. 2007. Variations in urine excretion of steroid hormones after an acute session
- and after a 4-week programme of strength training. Eur J Appl Physiol 99:65-71.
- 1430 [64] Tanner T, Marks R. 2008. Delivering drugs by the transdermal route: Review and
- comment. Skin Res Technol 14:249-260.
- 1432 [65] Chang H, Hu J, Shao B. 2007. Occurrence of natural and synthetic glucocorticoids in
- sewage treatment plants and receiving river waters. *Environ Sci Technol* 41:3462-3468.
- 1434 [66] Piram A, Salvador A, Gauvrit J-Y, Lanteri P, Faure R. 2008. Development and
- optimisation of a single extraction procedure for the LC/MS/MS analysis of two pharmaceutical
- classes residues in sewage treatment plant. *Talanta* 74:1463-1475.
- 1437 [67] Siemens J, Huschek G, Siebe C, Kaupenjohann M. 2008. Concentrations and mobility of
- human pharmaceuticals in the world's largest wastewater irrigation system, Mexico City-
- 1439 Mezquital Valley. Water Res 42:2124-2134.
- 1440 [68] OSPAR Commission. 2007. OSPAR List of Chemicals for Priority Action (Update 2007).
- 1441 Reference 2004-12. Convention for the Protection of the Marine Environment of the North-east
- 1442 Atlantic, OSPAR Commission on Hazardous Substances. London, UK.

- 1443 [69] Hilton MJ, Thomas KV, Ashton D. 2003. Targeted monitoring programme for
- pharmaceuticals in the aquatic environment. R&D Technical Report P6-012/06/TR. Environment
- 1445 Agency, Bristol, UK.
- 1446 [70] Amidon GL, Lennernas H, Shah VP, Crison JR. 1995. A theoretical basis for a
- biopharmaceutic drug classification: The correlation of in vitro drug product dissolution and in
- 1448 vivo bioavailability. *Pharm Res* 12:413-420.
- 1449 [71] Wu CY, Benet LZ. 2005. Predicting drug disposition via application of BCS:
- 1450 Transport/absorption/ elimination interplay and development of a biopharmaceutics drug
- disposition classification system. *Pharm Res* 22:11-23.
- 1452 [72] Custodio JM, Wu CY, Benet LZ. 2008. Predicting drug disposition,
- absorption/elimination/transporter interplay and the role of food on drug absorption. Adv Drug
- 1454 Delivery Rev 60:717-733.
- 1455 [73] Johnson R, Nusbaum BP, Horwitz SN, Frost P. 1983. Transfer of topically applied
- tetracycline in various vehicles. *Arch Dermatol* 119:660-663.
- 1457 [74] Kunz GJ, Klein KO, Clemons RD, Gottschalk ME, Jones KL. 2004. Virilization of young
- children after topical androgen use by their parents. *Pediatrics* 114:282-284.
- 1459 [75] Bhowmick SK, Ricke T, Rettig KR. 2007. Sexual precocity in a 16-month-old boy
- induced by indirect topical exposure to testosterone. Clin Pediatr (Phila) 46:540-543.
- 1461 [76] Franklin SL, Geffner ME. 2003. Precocious puberty secondary to topical testosterone
- 1462 exposure. J Pediatr Endocrinol Metab 16:107-110.
- 1463 [77] Yu YM, Punyasavatsu N, Elder D, D'Ercole AJ. 1999. Sexual development in a two-year-
- old boy induced by topical exposure to testosterone. *Pediatrics* 104:e23.

- This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).
- 1465 [78] Rolf C, Knie U, Lemmnitz G, Nieschlag E. 2002. Interpersonal testosterone transfer after
- topical application of a newly developed testosterone gel preparation. Clin Endocrinol 56:637-
- 1467 641.
- 1468 [79] ZumBrunnen TL, Meuwsen I, Michiel de V, Brennan JJ. 2006. The effect of washing and
- the absence of interindividual transfer of estradiol gel: A direct skin-to-skin contact study in
- postmenopausal women. American Journal of Drug Delivery 4:89-95.
- 1471 [80] Brachet C, Vermeulen J, Heinrichs C. 2005. Children's virilization and the use of a
- testosterone gel by their fathers. Eur J Pediatr 164:646-647.
- 1473 [81] Wester RC, Hui X, Maibach HI. 2006. In vivo human transfer of topical bioactive drug
- between Individuals: Estradiol. *J Investig Dermatol* 126:2190-2193.
- 1475 [82] Montgomery J. 2004. Strange progesterone saga. *Medical Laboratory Observer* 36:8.
- 1476 [83] Wolthuis A, de Vreeze J. 2005. Unexpected testosterone result for external quality
- assessment scheme sample. Clin Chem 51:475-476.
- 1478 [84] Richman C, Castensson S. 2008. Impact of waste pharmaceuticals: An environmental
- 1479 hazard or "greenwash"? *Pharm J* 280:335-342.
- 1480 [85] Marquardt KA, Tharratt RS, Musallam NA. 1995. Fentanyl remaining in a transdermal
- 1481 system following three days of continuous use. *Ann Pharmacother* 29:969-971.
- 1482 [86] McCormick CG. 2006. Regulatory challenges for new formulations of controlled
- substances in today's environment. *Drug Alcohol Depend* 83:S63-S67.
- 1484 [87] Evans JD, Evans EW. 2007. Review of eight restricted-access programs and potential
- implications for pharmacy. American Journal of Health-System Pharmacy 64:1302-1310.
- 1486 [88] Scheindlin S. 2004. Transdermal drug delivery: Past, present, future. *Mol Interv* 4:308-
- 1487 312.

- This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).
- 1488 [89] Musson SE, Townsend TG. 2009. Pharmaceutical compound content of municipal solid
- 1489 waste. *J Hazard Mater* 162:730-735.
- 1490 [90] Geurts MGJ, de Boer W, de Graaf JS, van Ginkel CG. 2007. Environmental exposure
- assessment of ethinyl estradiol (EE) from a combined hormonal vaginal contraceptive ring after
- disposal; leaching from landfills. *Sci Total Environ* 377:366-370.
- [91] Kidd KA, Blanchfield PJ, Mills KH, Palace VP, Evans RE, Lazorchak JM, Flick RW.
- 1494 2007. Collapse of a fish population after exposure to a synthetic estrogen. *Proc Nat Acad Sci*
- 1495 104:8897-8901.
- 1496 [92] Institute of Safe Medicine Practice. 2005. New fentanyl warnings: More needed to protect
- patients. European Journal of Hospital Pharmacy Practice 11:68.
- 1498 [93] Woodall KL, Martin TL, McLellan BA. 2008. Oral abuse of fentanyl patches (Duragesic):
- 1499 Seven case reports. J Forensic Sci 53:222-225.
- 1500 [94] Teske J, Weller JP, Larsch K, Troger HD, Karst M. 2007. Fatal outcome in a child after
- ingestion of a transdermal fentanyl patch. *Int J Leg Med* 121:147-151.
- 1502 [95] Horowitz R, Mazor SS, Aks SE, Leikin JB. 2005. Accidental clonidine patch ingestion in a
- child. *American Journal of Therapeutics* 12:272-274.
- 1504 [96] Tharp AM, Winecker RE, Winston DC. 2004. Fatal intravenous fentanyl abuse: four cases
- involving extraction of fentanyl from transdermal patches. Am J Forensic Med Pathol 25:178-
- 1506 181.
- 1507 [97] Martin TL, Woodall KL, McLellan BA. 2006. Fentanyl-related deaths in Ontario, Canada:
- Toxicological findings and circumstances of death in 112 cases (2002-2004). J Anal Toxicol
- 1509 30:603-610.

- 1510 [98] Woolf A, Burkhart K, Caraccio T, Litovitz T. 1997. Childhood poisoning involving
- transdermal nicotine patches. *Pediatrics* 99:e4.
- 1512 [99] Flannagan LM, Butts JD, Anderson WH. 1996. Fentanyl patches left on dead bodies --
- potential source of drug for abusers. *J Forensic Sci* 41:320-321.
- 1514 [100] Yerasi AB, Butts JD. 1997. Disposal of used fentanyl patches. Am J of Health Syst Pharm
- 1515 54:85-86.
- 1516 [101] Hamscher G, Pawelzick HT, Sczesny S, Nau H, Hartung J. 2003. Antibiotics in dust
- originating from a pig-fattening farm: A new source of health hazard for farmers? *Environ*
- 1518 *Health Perspect* 111:1590-1594.
- 1519 [102] U.S. Fish and Wildlife Service. 2008. SMARxT Disposal: A prescription for a healthy
- planet. The American Pharmacists Association and the Pharmaceutical Research and
- 1521 Manufacturers of America. Arlington, VA, USA.
- 1522 [103] De Bolle L, Mehuys E, Adriaens E, Remon J-P, Van Bortel L, Christiaens T. 2008. Home
- medication cabinets and self-medication: A Source of potential health threats? *Ann*
- 1524 *Pharmacother* 42:572-579.
- 1525 [104] Stubbs J, Haw C, Dickens G. 2008. Dose form modification a common but potentially
- 1526 hazardous practice. A literature review and study of medication administration to older
- psychiatric inpatients. *Int Psychogeriatr* 20:616-627.
- 1528 [105] Cone EJ. 2006. Ephemeral profiles of prescription drug and formulation tampering:
- Evolving pseudoscience on the Internet. *Drug Alcohol Dependence* 83:S31-S39.
- 1530 [106] Mitchell JF. 2009. Oral dosage forms that should not be crushed. Institute for Safe
- 1531 Medication Practices, Horsham, PA, USA.

- This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).
- 1532 [107] Pharmacist's Letter. 2007. Proper disposal of expired or unwanted drugs. *Pharmacist's*
- 1533 Letter 23:230401.
- 1534 [108] AB Innovations. 2008. Pill crushers. AB Innovations, Sumas, WA, USA.
- 1535 [109] Gebhart F. 2008. Pharmacy edges toward medication returns. *Drug Topics*. 152:25.
- 1536 [110] Halford B. 2008. Formulations for fighting abuse: Opioid makers aim to thwart misuse
- with excipients, additives, and antagonists. *Chem Eng News* 86:40-41.
- 1538 [111] Prüss A, Giroult E, Rushbrook P. 1999. Safe Management of Wastes from Health-care
- 1539 Activities: A Practical Guide. World Health Organization, Geneva, Switzerland.
- 1540 [112] Health Care Without Harm. August 2001. Non-incineration medical waste treatment
- technologies: A resource for hospital administrators, facility managers, health care professionals,
- environmental advocates, and community members. Arlington, VA, USA.
- 1543 [113] Shappell NW, Vrabel MA, Madsen PJ, Harrington G, Billey LO, Hakk H, Larsen GL,
- Beach ES, Horwitz CP, Ro K, Hunt PG, Collins TJ. 2008. Destruction of estrogens using fe-
- taml/peroxide catalysis. *Environ Sci Technol* 42:1296-1300.
- 1546 [114] Hirose J, Kondo F, Nakano T, Kobayashi T, Hiro N, Ando Y, Takenaka H, Sano K. 2005.
- 1547 Inactivation of antineoplastics in clinical wastewater by electrolysis. *Chemosphere* 60:1018-
- 1548 1024.
- 1549 [115] National Institute for Occupational Safety and Health. 2004. Preventing occupational
- exposure to antineoplastic and other hazardous drugs in health care settings, Appendix A: Drugs
- 1551 Considered Hazardous. Publication 2004-165. Atlanta, GA, USA.
- 1552 [116] Connor TH, McDiarmid MA. 2006. Preventing occupational exposures to antineoplastic
- drugs in health care settings. CA Cancer J Clin 56:354-365.

- This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).
- 1554 [117] American Society of Health-System Pharmacists. 2006. ASHP guidelines on handling
- hazardous drugs. *Am J Health-Syst Pharm* 63:1172-1193.
- 1556 [118] Meyer S, Eddleston M, Bailey B, Desel H, Gottschling S, Gortner L. 2007. Unintentional
- household poisoning in children. *Klin Pädiatr* 219:254-279.
- 1558 [119] Bar-Oz B, Levichek Z, Koren G. 2004. Medications that can be fatal for a toddler with one
- tablet or teaspoonful: A 2004 update. *Pediatric Drugs* 6:123-126.
- 1560 [120] Bradley TJ, Williams WH. 1975. Evaluation of medicines returned in Manchester DUMP
- 1561 campaign. *Pharm J* 215:542.
- 1562 [121] Harris DW, Karandikar DS, Spencer MG, Leach RH, Bower AC, Mander GA. 1979.
- Returned-medicines campaign in Birmingham. *Lancet* 313:599-601.
- 1564 [122] Doerr-MacEwen NA. 2007. The management of human pharmaceuticals in the
- environment. PhD thesis. University of Waterloo, Waterloo, ON, Canada.
- 1566 [123] Jjemba PK. 2006. Excretion and ecotoxicity of pharmaceutical and personal care products
- in the environment. *Ecotoxicol Environ Saf* 63:113-130.
- 1568 [124] Braybrook S, John DN, Leong K. 1999. A survey of why medicines are returned to
- 1569 pharmacies. *Pharm J* 263:7063.
- 1570 [125] Ruhoy IS. 2008. Examining unused pharmaceuticals in the environment. PhD thesis.
- 1571 University of Nevada, Las Vegas, NV, USA.
- 1572 [126] Boothby LA, Doering PL, Hatton RC. 2003. Carisoprodol: A marginally effective skeletal
- muscle relaxant with serious abuse potential. *Hospital Pharmacy* 38:337–345.
- 1574 [127] Soliman MA, Pedersen JA, Park H, Castaneda-Jimenez A, Stenstrom MK, Suffet IH.
- 1575 2007. Human pharmaceuticals, antioxidants, and plasticizers in wastewater treatment plant and
- water reclamation plant effluents. *Water Environ Res* 79:156-167.

- 1577 [128] Thomas Reid Associates. 2006. Santa Clara Valley Water District Stream Flow
- 1578 Augmentation Project: Upper Silver Creek and Coyote Creek Initial Study/Mitigated Negative
- Declaration. Santa Clara Valley Water District. San Jose, CA, USA.
- 1580 [129] Pedersen JA, Yeager MA, Suffet IH. 2002. Characterization and mass load estimates of
- organic, compounds in agricultural irrigation runoff. Water Sci Technol 45:103-110.
- 1582 [130] Gebhart F. 2008. Agencies flush original strategies in favor of promoting drug take-back
- programs. *Drug Topics* 152:25.
- 1584 [131] Lienert J, Burki T, Escher BI. 2007. Reducing micropollutants with source control:
- 1585 Substance flow analysis of 212 pharmaceuticals in faeces and urine. *Water Sci Technol* 56:87-96.
- 1586 [132] Johnson HL, Maibach HI. 1971. Drug excretion in human eccrine sweat. J Investig
- 1587 Dermatol 56:182-188.
- 1588 [133] Zlateva S, Marinov P, Sabeva Y. 2007. Determination of toxic substances in sweat secret
- of severe forms of poisoning Toxic coma. Clinical meaning. *Journal of IMAB* Book 1:83-85.
- 1590 [134] Barnes AJ, Smith ML, Kacinko SL, Schwilke EW, Cone EJ, Moolchan ET, Huestis MA.
- 1591 2008. Excretion of methamphetamine and amphetamine in human sweat following controlled
- oral methamphetamine administration. *Clin Chem* 54:172-180.
- 1593 [135] Huestis MA, Oyler JM, Cone EJ, Wstadik AT, Schoendorfer D, Joseph RE. 1999. Sweat
- testing for cocaine, codeine and metabolites by gas chromatography-mass spectrometry. *Journal*
- of Chromatography B: Biomedical Sciences and Applications 733:247-264.
- 1596 [136] Kidwell DA, Holland JC, Athanaselis S. 1998. Testing for drugs of abuse in saliva and
- sweat. Journal of Chromatography B: Biomedical Sciences and Applications 713:111-135.
- 1598 [137] Burns M, Baselt RC. 1995. Monitoring drug use with a sweat patch: An experiment with
- 1599 cocaine. *J Anal Toxicol* 19:41-48.

- This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).
- 1600 [138] Schwilke EW, Barnes AJ, Kacinko SL, Cone EJ, Moolchan ET, Huestis MA. 2006. Opioid
- disposition in human sweat after controlled oral codeine administration. Clin Chem 52:1539-
- 1602 1545.
- 1603 [139] Kintz P, Tracqui A, Jamey C, Mangin P. 1996. Detection of codeine and phenobarbital in
- sweat collected with a sweat patch. J Anal Toxicol 20:197-210.
- 1605 [140] Cirimele V, Kintz P, Gosselin O, Ludes B. 2000. Clozapine dose-concentration
- relationships in plasma, hair and sweat specimens of schizophrenic patients. Forensic Sci
- 1607 *Int*107:289-300.
- 1608 [141] Kintz P, Tracqui A, Mangin P. 1996. Sweat testing for benzodiazepines. J Forensic Sci
- 1609 41:851-854.
- 1610 [142] Faergemann J, Laufen H. 1993. Levels of fluconazole in serum, stratum corneum,
- epidermis-dermis (without stratum corneum) and eccrine sweat. Clin Exp Dermatol 18:102-106.
- 1612 [143] Shah VP, Epstein WL, Riegelman S. 1974. Role of sweat in accumulation of orally
- administered griseofulvin in skin. *J Clin Investig* 53:1673-1678.
- 1614 [144] Hoharitanon S, Chaichalotornkul J, Sindhupak W. 2005. A comparison of the efficacy
- between two itraconazole generic products and the innovative itraconazole in the treatment of
- 1616 tinea pedis. *J Med Assoc Thai* 88 (Suppl 4):S167-172.
- 1617 [145] Taylor JR, Watson ID, Tames FJ, Lowe D. 1998. Detection of drug use in a methadone
- maintenance clinic: Sweat patches versus urine testing. *Addiction* 93:847-853.
- 1619 [146] Fucci N, De Giovanni N, Scarlata S. 2008. Sweat testing in addicts under methadone
- treatment: An Italian experience. Forensic Sci Int 174:107-110.
- 1621 [147] Ishiyama I, Nagai T, Nagai T, Komuro E, Momose T, Akimori N. 1979. The significance
- of drug analysis of sweat in respect to rapid screening for drug abuse. *Int J Leg Med* 82:251-256.

- This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).
- 1623 [148] Huestis MA, Cone EJ, Wong CJ, Umbricht A, Preston KL. 2000. Monitoring opiate use in
- substance abuse treatment patients with sweat and urine drug testing. *J Anal Toxicol* 24:509-521.
- 1625 [149] Saito T, Wtsadik A, Scheidweiler KB, Fortner N, Takeichi S, Huestis MA. 2004.
- Validated gas chromatographic-Negative ion chemical ionization mass spectrometric method for
- 1627 Δ9-tetrahydrocannabinol in sweat patches. *Clin Chem* 50:2083-2090.
- 1628 [150] Huestis MA, Scheidweiler KB, Saito T, Fortner N, Abraham T, Gustafson RA, Smith ML.
- 1629 2008.4 Excretion of Δ9-tetrahydrocannabinol in sweat. *Forensic Sci Int* 174:173-177.
- 1630 [151] National Library of Medicine. 2009. DailyMed: Current Medication Information. U.S.
- National Library of Medicine and National Institutes of Health, Bethesda, MD, USA.
- 1632 [152] RxList. 2008. The Internet Drug Index. WebMD, New York, NY.
- 1633 [153] DrugLib.com. 2008. Drug Information Portal. Brooklyn, NY, USA.
- 1634 [154] Volmer DA, Hui JPM. 1998. Study of erythromycin A decomposition products in aqueous
- solution by solid-phase microextraction/liquid chromatography/tandem mass spectrometry.
- 1636 Rapid Commun Mass Spectrom 12:123-129.
- 1637 [155] Gulkowska A, He Y, So MK, Yeung LWY, Leung HW, Giesy JP, Lam PKS, Martin M,
- Richardson BJ. 2007. The occurrence of selected antibiotics in Hong Kong coastal waters. *Mar*
- 1639 *Pollut Bull* 54:1287-1293.
- 1640 [156] Batt AL, Bruce IB, Aga DS. 2006. Evaluating the vulnerability of surface waters to
- antibiotic contamination from varying wastewater treatment plant discharges. *Environ Pollut*
- 1642 142:295-302.
- 1643 [157] Spongberg AL, Witter JD. 2008. Pharmaceutical compounds in the wastewater process
- stream in Northwest Ohio. *Sci Total Environ* 397:148-157.

This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).

- 1645 [158] Christian T, Schneider RJ, Farber HA, Skutlarek D, Meyer MT, Goldbach HE. 2003.
- Determination of antibiotic residues in manure, soil, and surface waters. Acta Hydrochim
- 1647 *Hydrobiol* 31:36-44.
- 1648 [159] Thomas KV, Hilton MJ. 2004. The occurrence of selected human pharmaceutical
- 1649 compounds in UK estuaries. *Mar Pollut Bull* 49:436-444.
- 1650 [160] Roberts PH, Thomas KV. 2006. The occurrence of selected pharmaceuticals in wastewater
- effluent and surface waters of the lower Tyne catchment. Sci Total Environ 356:143-153.
- 1652 [161] Trenholm RA, Vanderford BJ, Drewes JE, Snyder SA. 2008. Determination of household
- 1653 chemicals using gas chromatography and liquid chromatography with tandem mass
- 1654 spectrometry. *J Chromatogr A* 1190:253-262.
- 1655 [162] Nakada N, Kiri K, Shinohara H, Harada A, Kuroda K, Takizawa S, Takada H. 2008.
- 1656 Evaluation of pharmaceuticals and personal care products as water-soluble molecular markers of
- 1657 sewage. *Environ Sci Technol* 42:6347-6353.
- 1658 [163] Nakada N, Tanishima T, Shinohara H, Kiri K, Takada H. 2006. Pharmaceutical chemicals
- and endocrine disrupters in municipal wastewater in Tokyo and their removal during activated
- 1660 sludge treatment. *Water Res* 40:3297-3303.
- 1661 [164] Nakada N, Shinohara H, Murata A, Kiri K, Managaki S, Sato N, Takada H. 2007.
- Removal of selected pharmaceuticals and personal care products (PPCPs) and endocrine-
- disrupting chemicals (EDCs) during sand filtration and ozonation at a municipal sewage
- 1664 treatment plant. *Water Res* 41:4373-4382.
- 1665 [165] Reeves AD, Patton D. 2005. Faecal sterols as indicators of sewage contamination in
- estuarine sediments of the Tay Estuary, Scotland: An extended baseline survey. *Hydrology and*
- 1667 Earth System Sciences 9:81-94.

- This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).
- 1668 [166] Kolpin DW, Skopec M, Meyer MT, Furlong ET, Zaugg SD. 2004. Urban contribution of
- pharmaceuticals and other organic wastewater contaminants to streams during differing flow
- 1670 conditions. Sci Total Environ 328:119-130.
- 1671 [167] Peng X, Yu Y, Tang C, Tan J, Huang Q, Wang Z. 2008. Occurrence of steroid estrogens,
- endocrine-disrupting phenols, and acid pharmaceutical residues in urban riverine water of the
- 1673 Pearl River Delta, South China. Sci Total Environ 397:158-166.
- 1674 [168] Verenitch SS, Lowe CJ, Mazumder A. 2006. Determination of acidic drugs and caffeine in
- municipal wastewaters and receiving waters by gas chromatography-ion trap tandem mass
- 1676 spectrometry. *J Chromatogr A* 1116:193-203.
- 1677 [169] Romano MJ, Dinh A. 2001. A 1000-fold overdose of clonidine caused by a compounding
- error in a 5-year-old child with attention-deficit/hyperactivity disorder. *Pediatrics* 108:471-472.
- 1679 [170] Tintinalli JE, Kelen GD, Stapczynski JS, Ma OJ, Cline DM, eds. 2003. Tintinalli's
- 1680 Emergency Medicine: A Comprehensive Study Guide, 6th ed. McGraw-Hill, New York, NY,
- 1681 USA.
- 1682 [171] Janssen Ortho. 2008. Ortho Evra (norelgestromin / ethinyl estradiol Transdermal System).
- 1683 10154403. Ortho-McNeil-Janssen-Pharmaceuticals, Manati, Puerto Rico.
- 1684 [172] Physician's Desk Reference. 2009. Medical Economics Data Production Company,
- 1685 Montvale, NJ, USA.
- 1686 [173] Friel PN, Hinchcliffe C, Wright JV. 2005. Hormone replacement with estradiol:
- 1687 conventional oral doses result in excessive exposure to estrone. Altern Med Rev 10:36-41.
- 1688 [174] Centers for Disease Control. 2003. Nicotine poisoning after ingestion of contaminated
- ground beef --- Michigan, 2003. Morbidity Mortality Weekly Report (MMWR) 52:413-416.

This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).

1690	[175] Key Pharmaceuticals. 2004. NITRO-DUR (nitroglycerin) - Transdermal Infusion System:
1691	Product Information. Kenilworth, NJ, USA.
1692	[176] Balit CR, Lynch A-M, Gilmore SP, Murray L, Isbister GK. 2006. Lignocaine and
1693	chlorhexidine toxicity in children resulting from mouth paint ingestion: A bottling problem. $J$
1694	Paediatr Child Health 42:350-353.
1695	[177] Sanofi-Aventis. 23 March 2006. Prescribing Information - Demerol (meperidine HCL) 50
1696	mg tablets. 104774. Bridgewater, NJ, USA.
1697	
1698	
1699	
1700	
1701	
1702	
1703	
1704	
1705	
1706	
1707	
1708	
1709	
1710	
1711	

This ET&C Paper in Press manuscript is in its original unedited form and has not been copyedited or formatted for final production. This manuscript is fully citable. ©2009 Society of Environmental Toxicology and Chemistry (SETAC).

1712 Figure Legend

1714

1713 FIGURE 1. PharmEcokinetics of Active Pharmaceutical Ingredients (APIs).

78

Table 1. Factors Determining the Significance of Disposal and Washing or Bathing in the Discharge of Active Pharmaceutical Ingredients (APIs) via Sewers <sup>a</sup>

Factor	Term	Importance to Contributing APIs	Relevant Information
Usage rate	UR	Total mass or moles of API consumed per time period (mass/time). Disposal of little-used medications contributes insignificantly to the overall combined levels of APIs in the environment, regardless of the portion disposed. This contrasts with disposal of small portions of medications that are used in large quantities.	Usage includes prescribed and (OTC) amounts that are:  (1) purchased in-country (including gray and black markets), (2) distributed for free (e.g., physician samples; community programs for indigents; charitable contributions from manufacturers), and (3) imported from other countries. One complication (discussed in the text) is that sales and disposal are not linked in time.  Disposal always occurs from sales made in the past. This time lag can also vary, forcing gross simplifications for the purposes of modeling.

Disposal	DP	Portion of total API left	One complication is the portion of leftover drugs that are
potential		over versus amount that	indefinitely stockpiled, never disposed. Also requiring
		was meant to be totally	disposal are used delivery systems still containing
		consumed as directed.	residual APIs (see: Type of delivery system or container,
			below). The potential for a drug to be disposed is
			probably partly a function of geographic locale and time
			of year (e.g., for seasonal medications).
Method of	PDs	Portion of API disposed to	How (and sometimes whether) a medication is disposed
disposal		sewers (flushed or poured	is partly a function of the design of the medication's
		down drains, such as in-	packaging. Some packaging (such as unit-packaged
		sink garbage disposals)	drugs) is more amenable to discarding to trash (because
		versus all other routes or	of the effort involved in removing from the package in
		fates (e.g., trash, burial,	order to flush), while other packaging is more likely to
		collection events,	result in disposal to sewers (e.g., bulk-packaged
		diversion, permanently	tablets/capsules and liquids); excess medication that
		stockpiled on site,	remains in delivery devices or delivery systems is also
		charitable contributions).	more prone to disposal via trash rather than sewers (see
		Note that charitable	Type of delivery or container, below), although some
		contribution is a route	delivery systems (such as patches) that contain potent
		generally relevant only in	APIs (which can cause unintentional poisonings) or those
		controlled healthcare	subject to abuse, may still require disposal via flushing.
		settings, e.g., physician	Charitable contributions (and drug sharing) merely
		donation of unexpired	postpone the eventual fate of APIs but might serve to
		samples.	reduce the need for new purchases.

Primary	PMP	Combined portions of	There are two extremes: (1) extensive metabolism, where
metabolic		systemic API excreted	little of the parent API [or glucuronides susceptible to
profile		unchanged into urine and	hydrolysis] is excreted, and (2) extensive excretion in
		feces and portion of	urine and feces, where the unchanged parent API is
		ingested dose not	excreted, sometimes stoichiometrically; some drugs, such
		absorbed by the gut. Data	as neomycin, are poorly absorbed after oral or parenteral
		come from	administration and therefore are excreted largely
		pharmacokinetics studies.	unchanged because of no opportunity for metabolism
		It is important to note,	(even if the small absorbed portion is extensively
		however, that excretion	metabolized). One complication is excretion of
		data can be highly variable	glucuronides having the potential to later be hydrolyzed
		for a given API and are	to products including the parent API; conjugates can
		often difficult to find in	therefore often be treated as parent API. Excretion
		the published literature	contributes primarily via urine and feces. For APIs that
		[131].	are extensively metabolized, alternate routes to sewers
			become comparatively more significant.
Secondary	SMP	Portion of systemic API	When excreted via sweat, APIs tend to be excreted
metabolic		excreted unchanged via	unchanged. The relative contributions from sweat
profile		sweat (and other minor	compared with urinary or fecal excretion is unknown and
		routes such as vomitus).	comparatively much smaller but measurable for many
			drugs. The end result is that this portion becomes washed
			from the skin or transferred by bodily contact to other
			surfaces (including people).

Route of Portion of API remaining RDMedications designed for topical use, such as gels, delivery on skin after topical creams, lotions, sprays, tinctures, ointments, plasters, (bioavailability) application (or delivery shampoos, foams, powders, and soaps, or for transdermal system). Whether the delivery (where only a fraction is actually absorbed medication is designed for across the skin and a portion is retained on the surface of topical use, where little the skin) are efficiently discharged to sewers as a result API is actually absorbed of their intended use, via bathing/washing. These dermally, and the majority medications contribute APIs to sewage as if they were remains on the skin. disposed directly. Transdermal patches also leave Medications designed for residual on the skin, residue that will later be washed external use efficiently away as if it had been applied topically. introduce APIs to sewage via bathing or washing (as well as laundering), as if they were disposed directly. These pharmaceuticals serve essentially as inputs tantamount to unintended disposal.

Type of delivery TDPortion of dose remaining Drugs administered via delivery devices (e.g., system or in delivery system and transdermal patches, vaginal rings) can retain very large container discharged to sewers. portions of their total APIs after use is completed because Delivery systems and only fractions of their contents are actually delivered containers often can have systemically (e.g., as little as 15% or less). Containers (e.g., injection vials) and injection/infusion devices can large amounts of residual APIs (some of which also contain residues. When disposed after use, these can remains inaccessible contribute substantial quantities of APIs. These used because of the design of devices need to be factored in with their unused new the system's device or counterparts as contributors during disposal. While most containers). These new or used devices are not disposed to sewers, some residues can then serve as must still be flushed according to the U.S. Office of used medications that then National Drug Control Policy (ONDCP) guidelines (one require disposal. example being fentanyl patches)<sup>b</sup>. Important to note is that the very drugs that are subject to waste minimization via charitable donations (as set up by various states) are the same ones that would otherwise most likely be disposed into the trash - not the sewer - simply because of the time and effort required in removing each dose from its packaging. It is most likely that the drugs currently disallowed for donations play the larger roles in disposal via the sewer.

WM (API mass discharged to sewers from disposal and washing) = UR(mass) • [(DP • PDs) + (SMP + RD + TD)] Sr (relative overall significance of secondary routes) = WM • [(UR - WM) • PMP]<sup>-1</sup>

<sup>&</sup>lt;sup>a</sup> The factors in this table can be used in the following two equations (described in the text):

<sup>&</sup>lt;sup>b</sup>([23], http://www.whitehousedrugpolicy.gov/publications/pdf/prescrip\_disposal.pdf)

Table 2. Overview of Active Pharmaceutical Ingredients (APIs) Measured in Sweat

API or class		
Chemical Abstracts	Concentration (on basis of volume of sweat)	D.C
Service Registry Number,	or Mass Collected on Sweat Patch or Wipe	Reference
freebase		
β-lactam antibiotics	Mean maximum concentrations: benzylpenicillin (axilla 2.6 μg/ml) ceftazidime (axilla, 28.4 μg/ml; forearm, 11 μg/ml) ceftriaxone (axilla, 8.9 μg/ml; forearm, 2.5 μg/ml) cefuroxime (axilla, 7.8 μg/ml) phenoxymethylpenicillin (axilla, 0.4 μg/ml)	[46]
aminopyrine (58-15-1) and antipyrine (60-80-0)	extensively excreted via sweat; up to 14 μg/ml after single 1-g oral doses	[132]
amitriptyline (50-48-6)	extensive excretion in sweat after acute poisoning (0.78-0.2 mg/L)	[133]
amphetamine (300-62-9)	median (range) after low and high doses: 15.5 (6.5-40.5) and 53.8 (34.0-83.4) ng/patch	[134]
carbamazepine (298-46-4)	present in sweat extensive excretion in sweat after acute poisoning (3.267 mg/L)	[33] [133]
clomipramine (303-49-1)	extensive excretion in sweat after acute poisoning (0.28 mg/L)	[133]
ciprofloxacin (85721-33-1)	2.2-5.5 μg/mL (during 7-day course of 750 mg/day oral dose)	[40]
cocaine (50-36-2)	33-3,579 ng/patch/30 min (from heat-induced sweating) 43-3,799 ng/wipe ng/mL: cocaine (378), benzoylecgonine (78.7), ecgonine methyl ester (74) up to 315 ng/patch (140 ng/L) during 1 week after nasal doses of 50-126 mg	[135] [136] [37] [137]
codeine (76-57-3)	11-1,123 ng/patch/30 min (from heat-induced sweating) 0-225 ng/patch/week 2-127 ng/patch/day (after 90-mg oral dose)	[135] [138] [139]
clomipramine (303-49-1)	extensive excretion in sweat after acute poisoning: (0.28 mg/L)	[133]
clozapine (5786-21-0)	49 to 5,609 ng/patch in sweat after oral dosing of 200-700 mg/day	[140]
diazepam (439-14-5)	0.1-6 ng/patch total after one dose (first detected 2-4 h after dose; also detected was nordiazepam but not oxazepam) extensive excretion in sweat after acute poisoning	[141] [133]
doxorubicin (23214-92-8)	observed	[47, 48]
` '		- ' -

	·	I		
fentanyl (437-38-7)	concentrations in sweat varied from 0.17 to 1.02 ng/µl	[41]		
fluconazole (86386-73-4)	high concentrations in sweat, all above the serum concentrations	[142]		
griseofulvin (126-07-8)	200-300 ng/mL independent of sweat volume; after 0.5-g oral doses at 12-h intervals.	[143]		
itraconazole (84625-61-6)	mostly excreted through sebaceous glands; moderately excreted by the sweat glands	[144]		
loratadine (79794-75-5)	detectable on skin 40 min after ingesting a 10-mg oral dose	[44]		
MDMA ("ecstasy") (3,4- methylenedioxy- methamphetamine) (42542-10-9)	Mean: 542 ng/patch/day; range: 42.4-1,326 ng/patch/day accumulated after single 100-mg oral dose (first observed 1.5 h after dose)			
methadone (76-99-3)	presence correlated with urine detected in sweat of heroin addicts undergoing treatment			
Methamphetamine-HCl (51-57-0)	median (range) after low and high doses: 63.0 (16.8-175) and 307 (199-607) ng/patch constant rate of 1.4 μg/mL after oral dose of 10 mg			
methotrexate (59-05-2)	Mean 725 ng/mL (mean maximal concentration 1.7 μg/mL)	[50]		
median concentrations (ng/mL): heroin (10.5), 6-acetylmorphine (13.6), morphine (15.9), and codeine (13.0)		[148]		
phenobarbital (50-06-6) (phenobarbitone)	6-6) 0.5-33 ng/patch/day (first observed 3 h after 100-mg oral dose) concentration in sweat found to increase with increasing sweat flow			
phenytoin (57-41-0)	concentration in sweat was independent of sweat flow			
sulfonamides	one of the very early studies documenting that drugs are excreted via sweat; sulfapyridine, sulfathiazole, sulfadiazine, and <i>p</i> -aminohippurate ranged up into the 10's of µg/ml			
etrahydrocannabinol 0.9-3.11 ng/patch/day below detection limit after daily ingestion of 14.8 mg				

# Table 3. Active Pharmaceutical Ingredients (APIs) Commonly Used in Topical Medications<sup>a</sup>

### **STEROIDS**

Alclometasone dipropionate <sup>b</sup> 0.05%; cream, ointment Ameinonide 0.1%; cream, ointment, lotion

Betamethasone dipropionate <sup>d</sup> 0.05%; ointment, cream, lotion, gel

Betamethasone valerate d 0.12%; foam, lotion, cream

Clobetasol propionate 0.05%; foam, lotion, cream, ointment, gel, shampoo, spray

Clocortolone pivalate 0.1%; cream

Desonide 0.05%; foam, ointment, lotion, gel, powder, aerosol

Desoximetasone <sup>b</sup> 0.25%; cream, ointment, gel

Diflorasone diacetate 0.05%; cream, ointment

Fluocinolone acetonide 0.025%; cream, oil (also vitreal implant)

Fluocinonide 0.05%; cream, ointment, gel, solution

Flurandrenolide 4 µg/cm<sup>2</sup>; tape

Flurandrenolide 0.05%; ointment, cream, lotion

Fluticasone propionate <sup>c</sup> 0.05%; cream, lotion, ointment, spray

Halcinonide 0.1%; cream, ointment, solution

Halobetasol propionate 0.05%; cream, ointment

Hydrocortisone acetate <sup>c</sup> 2.5% (with pramoxine HCl 1%); lotion, cream, ointment

Hydrocortisone butyrate c 0.1%; cream, ointment

Hydrocortisone (cortisol) <sup>c</sup> 1%, iodoquinol 1%; cream

Hydrocortisone valerate c 0.2%; cream, ointment

Mometasone furoate <sup>b</sup> 0.1%; cream, ointment, lotion, spray

Prednicarbate 0.1%; cream, ointment

Triamcinolone acetonide c 0.2%; cream, lotion, ointment, aerosol

## **ACNE**

Adapalene 0.1%, 0.3%; gel

Clindamycin phosphate d 1.2%; gel, cream, foam, lotion, pads Erythromycin d 5%; gel, solution, ointment, swabs

Sulfacetamide, sodium 10%; lotion, ointment, cream, foam, gel, wash, pads

Tazarotene 0.1%; gel, cream

e Tretinoin 0.1%; alcohol, gel, cream, solution

## SKIN/EYE INFECTIONS

<sup>e</sup> Acyclovir 5%; cream, ointment, solution

Bacitracin zinc 500 Units; ointment

Butenafine HCl b 1%; cream

Chloroxine 2%; shampoo

Ciclopirox<sup>b, g</sup> 1%; shampoo, cream, lotion, gel

Ciclopirox <sup>b</sup> 8%; topical solution (nail lacquer)

Clotrimazole c 1%; cream, lotion, solution

Docosanol 10%: cream

Econazole nitrate 1%; cream

Ketoconazole c 2%; gel, shampoo, foam, cream

Miconazole nitrate 2%; powder, spray, cream, suppositories

Mupirocin 2%; ointment, cream

Naftifine HCl<sup>b</sup> 1%; cream, gel

e Neomycin 3.5 mg (with bacitracin zinc 500 Units/g, polymyxin B sulfate 10,000 Units/g); ointment

e Nystatin 100000 Units/g; powder

Oxiconazole nitrate 1%; cream, lotion

Penciclovir 1%; cream

e Polymyxin B sulfate 10000 Units/g (with bacitracin zinc 500 Units/g or neomycin sulfate 0.35%); ointment, powder, cream

Retapamulin 1%; ointment

Sertaconazole nitrate 2%; cream

Sulconazole nitrate 1%; cream, solution

<sup>e</sup> Sulfadiazine, silver 1%; cream

Terbinafine HCl<sup>c</sup> 1%; cream, solution

<sup>e</sup> Tobramycin 0.3% ointment, solution

Tolnaftate 1%; cream, powder, solution, aerosol

#### PSORIASIS

Calcipotriene <sup>b</sup> 0.005%; ointment, solution, cream Salicylic acid 6%; shampoo

#### WARTS

Imiquimod <sup>b</sup> 5%; cream Podofilox <sup>b</sup> 0.5%; gel, solution Salicylic acid 40%; plaster

### SCABIES/LICE

Crotamiton 10%; cream, lotion

Lindane b 1%; lotion, shampoo

Malathion <sup>b</sup> 0.5%; lotion

Permethrin 5%; cream

Pyrethrins 0.33%, piperonyl butoxide 4%; gel, shampoo, lotion

Pyrethrum extract 0.33%, piperonyl butoxide 4%; oil

## **ROSACEA**

Azelaic acid b 20%; cream, gel

<sup>e</sup> Metronidazole 1%; lotion, cream, gel

### LOCAL ANESTHETICS

Benzocaine, Butamben, Dibucaine, Lidocaine, Pramoxine, Tetracaine; cream, ointment, gel, lotion

## MISCELLANEOUS

e Doxepin HCl 5%; cream

<sup>e</sup> Fluorouracil 5%; solution, cream

Nitroglycerin <sup>c</sup> 2%; ointment, solution, patch

Pimecrolimus b 1%; cream

Tacrolimus d 0.03%; ointment

- <sup>a</sup> This listing ([57], http://formulary.prescribingreference.com/dermatological\_disorders) excludes those medications that are specially formulated as transdermal systems (e.g., patches).
- NOTE: Only the highest concentration in use is listed, which may not apply to all the formulations. APIs used in multiple categories are listed only under the category that uses the highest concentration. Usage and excretion data from: ([151], http://dailymed.nlm.nih.gov/dailymed/about.cfm; [152], http://www.rxlist.com; [153], http://www.druglib.com).
- Unless otherwise noted, all APIs are approved only for external use (topical, dermatologic, ophthalmic), having no common off-label oral or parenteral use.
- <sup>b</sup> API is used only externally, but a small portion is known to be systemically absorbed and excreted.
- <sup>c</sup> API also has oral and/or parenteral use but is extensively metabolized and therefore little is excreted unchanged.
- <sup>d</sup> API also has oral and/or parenteral use and a small portion (<10%) is excreted unchanged.
- <sup>e</sup> API also has oral and/or parenteral use and is extensively excreted unchanged or as active metabolites.
- <sup>a,f</sup> Erythromycin readily undergoes internal dehydration to inactive anhydroerythromycin (erythromycin-H<sub>2</sub>O) [154], which is routinely detected in the environment [155].
- b,g Approximately 10% of ciclopirox dermal dose excreted unchanged over 10 hours ([153], http://www.druglib.com).

Table 4. Topical Active Pharmaceutical Ingredients (APIs) Poorly Excreted but Identified During Environmental Monitoring<sup>a</sup>

API Chemical Abstracts Service Registry Number	Sewage Influent ng/l	Sewage Effluent ng/l	Surface Water ng/l	Biosolids ng/kg dry weight	Reference
Betamethasone (378-44-9) (plus dexamethasone; the two could not be distinguished)	15	7	0.02-0.31		[66] [65]
Clindamycin (18323-44-9)	80-120	1,000 30	up to 24	1,540	[156] [157] [158] [67]
Clotrimazole (23593-75-1)	10-33		up to 22 6-34		[159] [160]
Cortisone (53-06-5)  Cortisol (hydrocortisone) (50-23-7)	174 4.6-86 53 7.6-120 370	229 0.13-0.58 63 0.25-1.9 38	0.06-4.2 0.08-3.4		[66] [65] [66] [65] [161]
Crotamiton (483-63-6)	1,610	245-968 580-979	up to 504 6.67 (groundwater) 269-504		[162] [163] [164]

Docosanol (661-19-8)			reported in river sediments (but source might be natural)		[165]
Fluocinolone acetonide (67-73-2)	0.3	11			[66]
Miconazole (22916-47-8)			to 9		[160] [166]
Salicylic acid (69-72-7)		25-47 up to 2,100	up to 2,100 370	96,000-253,000	[157] [167] [168]
Triamcinolone acetonide (76-25-5)	40	3			[66]

<sup>&</sup>lt;sup>a</sup>APIs from Table 3, for which the potential is highest for dermal application (as opposed to excretion) as a source for environmental residues; all of the scabies/lice APIs have been excluded because of their large non-therapeutic usages in pest control.

Table 5. Active Pharmaceutical Ingredients (APIs) Commonly Used in Delivery Devices for Administering Sustained Dermal Doses<sup>a</sup>

API – device (trade name) Chemical Abstracts Service Registry Number freebase	Mass/Device	Daily oral dose (if applicable); unless otherwise noted	Lethal dose	Equivalent lethal oral doses per device	Excretion Efficiency <sup>b</sup>	Equivalent oral daily doses contributing API to environment if device is disposed to sewers (range) c
fentanyl - transdermal patch (pain) - UNUSED [Duragesic] (437-38-7)	1.25-10 mg/patch	200 μg/day up to 1,200	~ 1 mg [85]	1-10 (for 70 kg person)	>90% transformed to N-dealkylated and hydroxylated	1.25 mg/(1.2 • 0.1) = 10 10 mg/(0.2 • 0.1) = 500
fentanyl - transdermal patch (pain) - USED	reported to retain 28-84% of original fentanyl content [85]	μg/day oral		<9 (for 70 kg person)	inactive metabolites	$(1.25 \cdot 0.28)/(1.2 \cdot 0.1) = 3$ $(10 \cdot 0.84)/(0.2 \cdot 0.1) = 420$
clonidine - transdermal patch [Catapres-TTS] ( 4205-90-7)	2.5-7.5 mg/patch per week	0.1-0.3 mg/day oral	highly toxic with ingestion by child of 0.01-0.04 mg/kg [169]	ca.10-30 (20-kg child)	40-60% excreted unchanged	2.5 mg/0.3 • 0.6 = 14 7.5 mg/0.1 • 0.4 = 188
methylphenidate - transdermal system [Daytrana] (113-45-1)	27-82 mg/patch	2.5-10 mg/day oral; max 90 mg/day	Unknown (children) 2-5 g (adults)	0	Only small quantities (<1%) of unchanged methylphenidate appear in the urine. Most of dose is excreted in urine as ritalinic acid (60%-86%), the remainder comprising minor metabolites	27 mg/90 • 0.01 = 30 82 mg/2.5 • 0.01 = 3,280
selegiline - transdermal system (depression) [Emsam] (14611-51-9)	20-40 mg/patch	10 mg/day oral	>140 mg [170]	0	extensively metabolized	20 mg/10 • 0.1 = 20 40 mg/10 • 0.1 = 40
rotigotine - transdermal system (Parkinson's) [Neupro] (99755-59-6)	4.5-13.5 mg/patch	NA: not used orally - high clearance and a relatively short duration of effect	> 0.1 mg/ml plasma concentration	0	extensively metabolized	Unlimited
rivastigmine - patch (Exelon) - reversible cholinesterase inhibitor (Alzheimer's; Parkinsons) [Exelon] (123441-03-2)	9-18 mg/patch	1.5-6.0 mg twice a day			extensively metabolized; no parent drug detectable	9 mg/6 • 0.01 = 17 18 mg/1.5 • 0.01 = 1,200

oxybutynin - transdermal (antispasmodic, anticholinergic) [Oxytrol] (5633-20-5)	36 mg/patch	5-15 mg/day oral		extensively metabolized; <0.1% excreted unchanged in urine	36 mg/15 • 0.01 = 240 36 mg/5 • 0.01 = 720
ethynylestradiol (EE2) (57-63-6) with norelgestromin (NGMN) patch (53016-31-2) (contraception) [Ortho-Evra]	6.00 mg NGMN 0.75 mg EE2/patch (per week)	250 μg NGMN/day (1.75 mg/week) 35 μg EE2/day (0.245 mg/week) Female hypogonadism: 0.02-0.05 mg EE2 1-3 times daily for first 2 weeks of cycle. Inoperable progressing prostatic cancer: from three 0.05 mg to four 0.5 mg daily for palliation. Inoperable progressing breast cancer: two 0.5 mg three times daily.		EE2: very low but can undergo extensive deconjugation  NGMN: extensively metabolized, but also to active metabolites ([171], http://www.orthoevra.com/orthoevra/shared/shared/pi/OrthoEvra PI.pdf)	EE2: 0.75 mg/0.035 • 0.1 = 214 NGMN: 6 mg/0.250 • 0.1 = 240
17β-estradiol (E2) - transdermal system [Estraderm; Menostar; Esclim; Alora; Vivelle; Climara] (50-28-2)	0.39-1.56 mg/patch	Replacement therapy oral doses ~ 1-2 mg/day (up to 30 mg/day for breast cancer) d		10% of oral dose excreted unchanged in urine [173], but can undergo extensive deconjugation; urinary excretion of endogenous estradiol ranges	$0.39 \text{ mg/2} \bullet 0.1 = 2$ $1.56 \text{ mg/2} \bullet 0.1 = 8$
17β-estradiol - metered- dose transdermal spray (EvaMist)	each metered dose (1.7%) contains 1.53 mg E2 (1-3 doses per day)	Replacement therapy oral doses ~ 1-2 mg/day (up to 30 mg/day for breast cancer)		5-100 µg/day (women), 2-25 µg/day (men), but up to 30 mg/day (pregnant women) [60]	1.53 mg/2 • 0.1 = 8
E2/levonorgestrel (797-63-7) - transdermal system [Climara Pro]	4.4 mg E2 and 1.39 mg levonorgestrel	E2: ~ 1-2 mg/day levonorgestrel: 0.1 mg/day		levonorgestrel partly excreted unchanged in urine	E2: 4.4 mg/2 • 0.1 = 22 4.4 mg/1 • 0.1 = 44 levonorgestrel: 1.39 mg/0.1 • 0.5 = 28
E2/norethindrone acetate (NETA) (51-98-9) - transdermal system [Combipatch]	0.62 -0.51 mg E2 and 2.7 - 4.8 mg NETA/patch	0.5 mg of norethindrone		<5% excreted unchanged, but extensively conjugated	NETA: 2.7 mg/0.5 • 0.05 = 108 4.8 mg/0.5 • 0.05 = 192

testosterone - transdermal system [Androderm] (58-22-0)	12-24 mg/patch	50-400 mg (testosterone enanthate, IM) once or twice per month		0	testosterone not usually administered orally (excretion of free and conjugated testosterone from adult males ranges up to 0.3 mg/day calculated from data in [62,63] About 90% given intramuscularly excreted as urine conjugates, and about 6% excreted unchanged in feces.	equiv oral DD = 2.5-5 mg/day (assuming no metabolism) $12 \text{ mg/}400 \bullet 0.1 = <1$ $24 \text{ mg/}50 \bullet 0.1 = 5$
diclofenac epolamine - topical patch (pain) [Flector] (119623-66-4)	180 mg/patch	100-200 mg/day oral, of diclofenac HCl		0	little excreted unchanged in urine	180 mg/200 • 0.1 = 9 180 mg/100 • 0.1 = 18
nicotine [Nicotrol; Nicoderm CQ] (54-11-5)	7-21 mg/patch	NA	30-60 mg [174]	<1 (toxic for children)	10-30% excreted unchanged in urine	21 mg/30 • 0.3 = 2
scopolamine - patch (nausea) [Transderm SCOP] (51-34-3)	1.5 mg/patch	oral dose 0.4 mg every 4-8 hours as needed	>2-4 mg	<1	<10% excreted unchanged in urine	1.5 mg/0.4 • 0.1 = 38
flurandrenolide - topical tape (corticosteroid) (1524-88-5)	4 mg/square centimeter	NA				Unlimited
nitroglycerin (angina) [Minitran; Nitro-Dur] (55-63-0)	20-160 mg/patch	~ 3 X 0.6 mg sublingual	>200-1,200 mg <sup>e</sup>	<1	extensively metabolized	20 mg/3 • 0.1 = 67 160 mg/0.6 • 0.1 = 2,667
lidocaine [lignocaine] - patch (5%) [Lidoderm] (137-58-6)	700 mg/patch; 46 mg/mucoadhesive patch	NA	Severe effects >15mg/kg [176]	>2 (20-kg child)	lidocaine: <10% excreted unchanged, but several less- potent metabolites are also	NA
lidocaine/tetracaine (94-24-6) [Synera]	70 mg each/patch	NA			excreted; tetracaine: unknown but undergoes rapid hydrolysis	NA
salicylic acid (warts) [Duofilm; Duoplant] (69-72-7)	17% flexible collodion	NA				NA

2009. Medical Economics Data Production Company, Montvale, NJ, USA.)

- c (mass contained in device) [(dose in mass per day) (fraction of API excreted unchanged)]<sup>-1</sup>; calculated to provide the minimum and maximum possible.
- <sup>d</sup> ([172], Physician's Desk Reference. 2009. Medical Economics Data Production Company, Montvale, NJ, USA.)

<sup>&</sup>lt;sup>a</sup> unless noted otherwise, data from: ([152], http://www.rxlist.com; [153], http://www.druglib.com; [172], Physician's Desk Reference.

<sup>&</sup>lt;sup>b</sup> extensive metabolism was assumed to equate with 10% of dose excreted unchanged;

<sup>&</sup>lt;sup>e</sup> ([175], STET http://www.spfiles.com/pinitrodur.pdf).

page 16 of 19

Table 6. Unknowns/Variables in Calculating Relative Environmental Footprint of a Disposed Drug

total consumption	No ready source of data on total quantity of an API consumed (on the basis of either locale or population). Must be derived from sales figures and by making assumptions of average cost per dose and average mass per dose. When using local coroner data [15], this can be derived from the dispensed amounts (but then it must also be known whether the medication was for short-term treatment or long-term maintenance). Multiple medications sharing the same API further complicate calculations.
fraction disposed	Even though coroner data specify the route of disposal, and even if the fraction disposed were known for the larger population, assumptions would still be required as to what portion was disposed via sewage versus trash or take-backs (or stockpiled indefinitely). One complication (discussed in the text) is that sales and disposal are not linked in time. Disposal always occurs from sales made in the past. This time lag can also vary, forcing gross simplifications for the purposes of modeling.
fraction excreted unchanged	Pharmacokinetic data are generally available for nearly all APIs <sup>a</sup> , but there are three major caveats: (1) these data are usually acquired from healthy volunteers, and absorption, metabolism, and excretion could differ widely for patients in diseased states or because of gender, ethnicity, age, body weight, diet, and other factors; (2) the metabolism of many drugs yields conjugates, which can be hydrolyzed during sewage treatment or once in the environment, yielding the parent API; and (3) for drugs administered with a delivery device (e.g., patches), the amount of API remaining in the used or partially used device is unknown, and this residue comprises large amounts of fully unmetabolized parent API.

<sup>&</sup>lt;sup>a</sup> ([152], http://www.rxlist.com; [153], http://www.druglib.com; [172], Physician's Desk

Reference. 2009. Medical Economics Data Production Company, Montvale, NJ, USA.)

Table 7. Drugs That Should Be Disposed by Flushing<sup>a</sup>

Drug Tradename Active Pharmaceutical Ingredients (API) [Chemical Abstracts Service Registry Number freebase]	Excretion Efficiency b	API Disposed (mg) <sup>c</sup>
dActiq (oral transmucosal fentanyl citrate) [990-73-8]	>90% transformed to N-dealkylated and hydroxylated inactive metabolites	0
dAndroGel (1% testosterone gel) [58-22-0]	testosterone not usually administered orally [excretion of free and conjugated endogenous testosterone from adult males ranges up to 0.3 mg/day calculated from data in [62,63] ~ 90% given intramuscularly excreted in urine as conjugates, and about 6% excreted unchanged in feces.]	4,430 total (16 x 5-mg patches = 80 mg 3 x 75-g 1% pumps = 2,250 mg 42 x 5-g 1% packets = 2,100 mg)
dAvinza (morphine sulfate extended-release) [64-31-3]	Approximately 10% of morphine dose excreted unchanged in the urine; 7-10% excreted in feces; most excreted as conjugates; but also excreted as the major metabolite of codeine and heroin	103,000
Baraclude (entecavir) [142217-69-4]	predominantly eliminated in urine as unchanged API (62-73% of dose)	0
<sup>d</sup> Daytrana (methylphenidate) [113-45-1]	Only small quantities (<1%) of unchanged methylphenidate appear in the urine. Most of the dose is excreted in the urine as ritalinic acid (60%-86%), the remainder being accounted for by minor metabolites	0
<sup>d</sup> Demerol (meperidine) [57-42-1]	negligible excretion unchanged ([177], http://www.sanofi-aventis.ca/products/en/demerol.pdf)	81,000
dDiastat AcuDial (diazepam rectal gel) [439-14-5]	well absorbed following rectal administration (equivalent of 90% of oral dose); extensively metabolized to conjugates	0
<sup>d</sup> Dilaudid/Dilaudid-HP (hydromorphone) [466-99-9]	extensively metabolized (>95%)	5,870
<sup>d</sup> Dolophine (methadone) [76-99-3]	extensively metabolized	53,480
dDuragesic (new and used) (fentanyl) [437-38-7]	see Actiq	2.9 total (3 x 25-µg patches 38 x 75-µg patches)

dEstrogel (estradiol gel; 0.06%) [a 1.25-g dose contains 750 µg; a new 93-g dispensing pump contains 55 mg; a fully used pump will retain about 10% residual] [50-28-2]	10% of oral dose excreted unchanged in urine [173], but can undergo extensive deconjugation; urinary excretion of endogenous estradiol ranges 5-100 μg/day (women), 2-25 μg/day (men), but up to 30 mg/day (pregnant women) [60]; replacement therapy oral doses < 1mg/day	amount washed from skin and hands (after application/absorption); dermal absorption efficiency is about 17% per day [61]
dFentora (fentanyl buccal tablets) [437-38-7]	see Actiq	0
<sup>d</sup> Ionsys (transdermal fentanyl)	see Actiq	0
Opana/Opana ER (oxymorphone) [76-41-5]	poorly absorbed; 50% excreted unchanged in urine	0
<sup>d</sup> OxyContin (oxycodone) [76-42-6]	4% excreted unchanged	271,636
<sup>d</sup> Percocet (oxycodone)		
dReyataz (atazanavir) [198904-31-3]	7% unchanged in urine	14,000
Suboxone (buprenorphine/naloxone)	great variability in excretion of buprenorphine; mainly excreted in feces; conjugates excreted in urine	0
Subutex (buprenorphine) [52485-79-7]		0
Tequin (gatifloxacin) [112811-59-3]	70% unchanged in urine	12,500
Tyzeka (telbivudine) [3424-98-4]	extensively excreted unchanged	0
<sup>d</sup> Videx/Videx EC (didanosine) [69655-05-6]	extensively metabolized	0
<sup>d</sup> Xyrem (sodium oxybate) [502-85-2]	<5% excreted unchanged in urine	0
Zerit (stavudine) [3056-17-5]	16-62% unchanged in urine	2,120

- <sup>a</sup> Disposal by flushing recommended by OND.
- ([23] http://www.whitehousedrugpolicy.gov/publications/pdf/prescrip\_disposal.pdf) and/or manufacturers.)
- <sup>b</sup> Note, however, that this assessment ignores the possible contributions from hydrolyzable conjugates or bioactive metabolites. Also note that elimination data for APIs is derived from testing on healthy humans. The actual percentage of clearance of unchanged API could be lower (but probably higher) in diseased patients or from those with certain metabolic polymorphisms.
- <sup>c</sup> Data acquired from a single county (Clark County, NV, USA) over the course of 12 months [15].
- d Drugs for which disposal may play a more dominant role in contributing to environmental residues; unless otherwise noted, pharmacokinetic data compiled from: ([152], http://www.rxlist.com;
   [153], http://www.druglib.com; [172], Physician's Desk Reference.2009. Medical Economics Data
   Production Company, Montvale, NJ, USA.

